

# Soft x-ray microscopy

## (Zone-plate based soft-x-ray spectro-microscopy)

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\* zone plate as a soft x-ray lens & probing depth

## 2. Types of microscopy and application examples

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2.2. Scanning transmission x-ray microscopy (STXM)

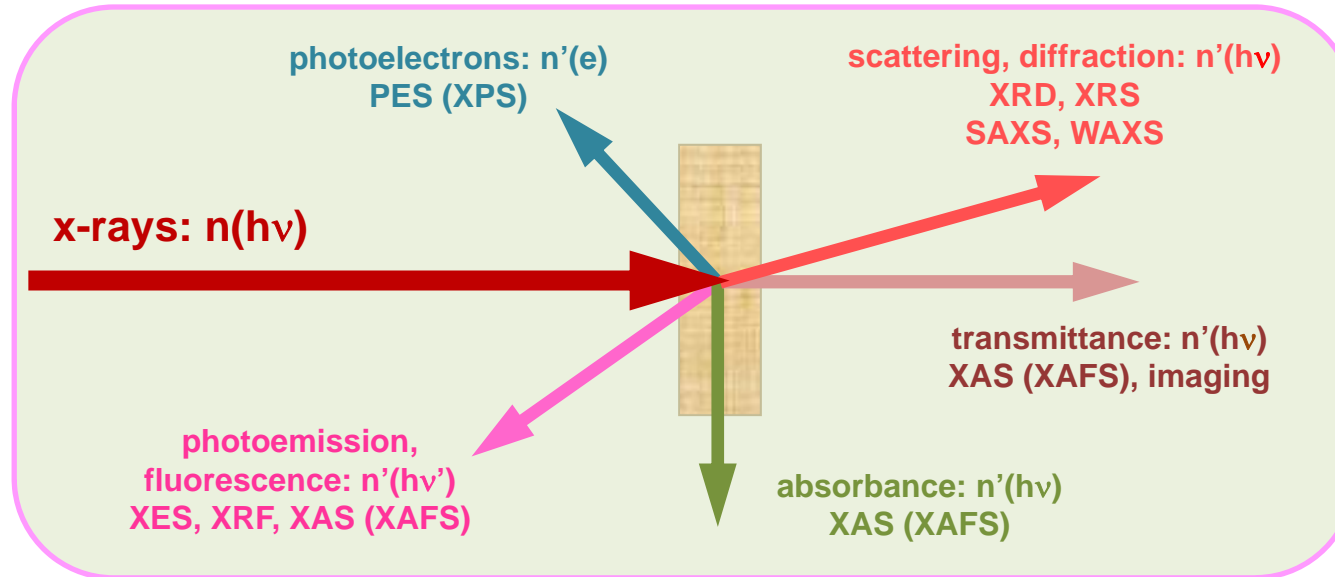
- Ptychography mode

- Fluorescence mode

2.3. Scanning photoelectron microscopy (SPEM)

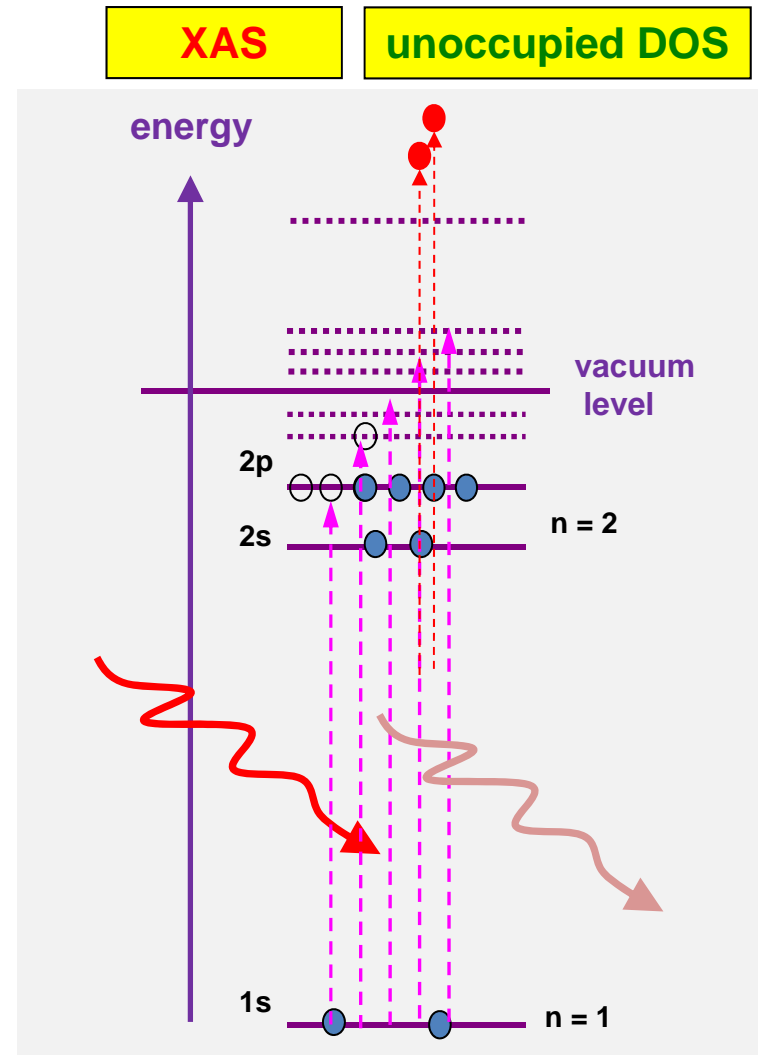
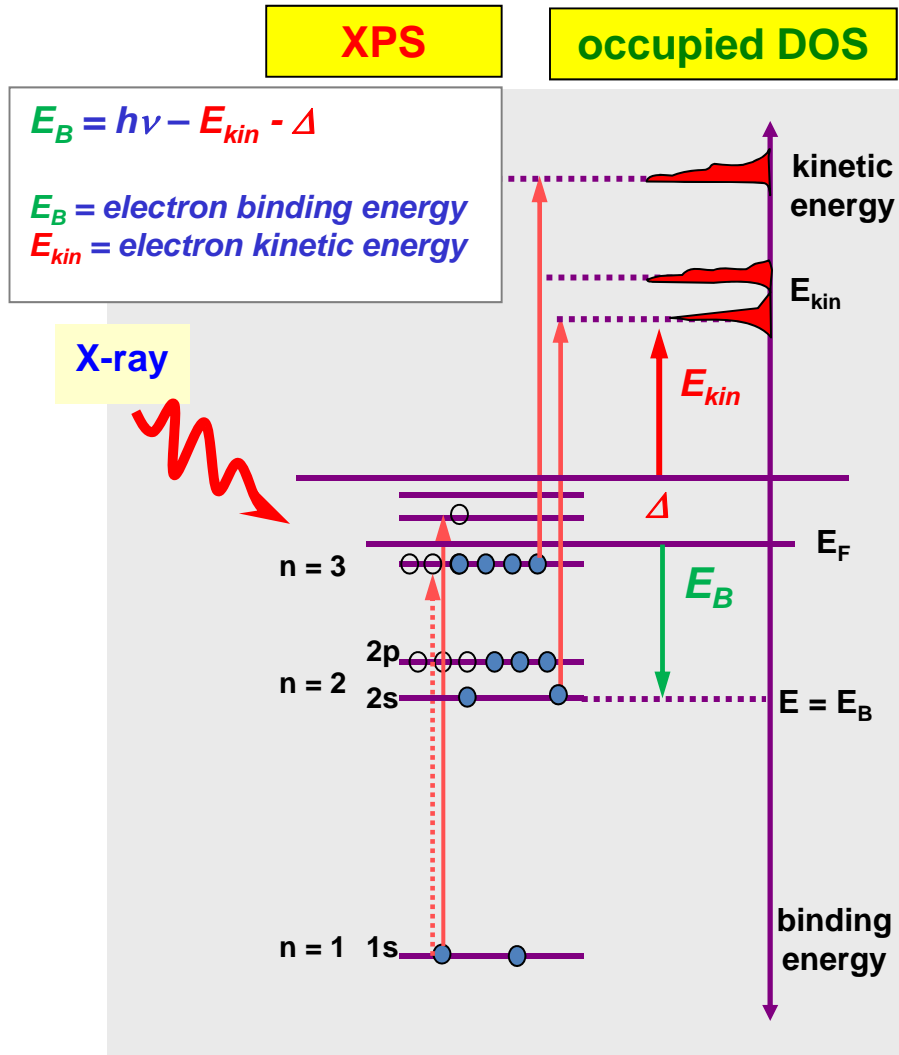
## 3. Summary

# 1. Use of x-rays



**PES: photoemission/ photoelectron spectroscopy**  
**XPS: X-ray photoemission/ photoelectron spectroscopy**  
**XRD: X-ray diffraction**  
**XRS: X-ray scattering**  
**SAXS: Small angle x-ray scattering**  
**WAXS: Wide angle x-ray scattering**  
**XAS: X-ray absorption**  
**XAFS: X-ray absorption fine structure**  
**XES: X-ray emission spectroscopy**  
**XRF: X-ray fluorescence**

1. XPS & XAS for materials science...

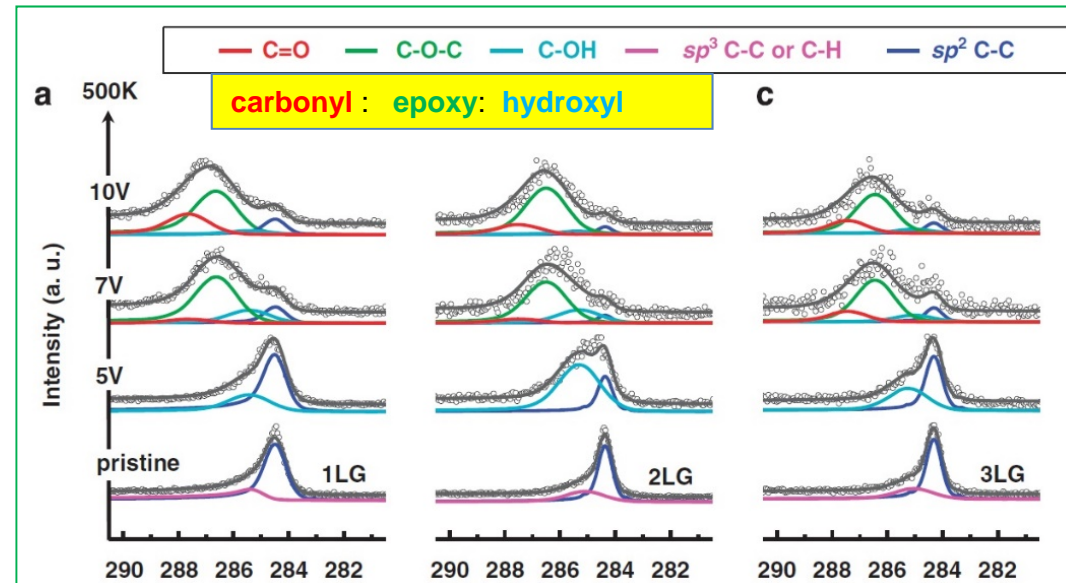
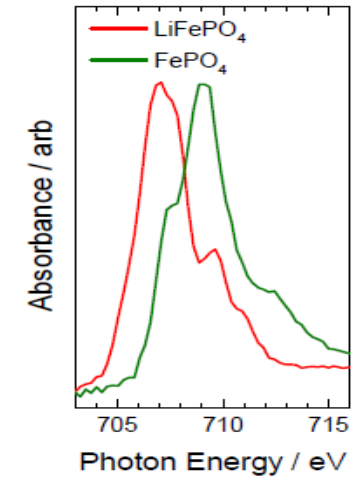
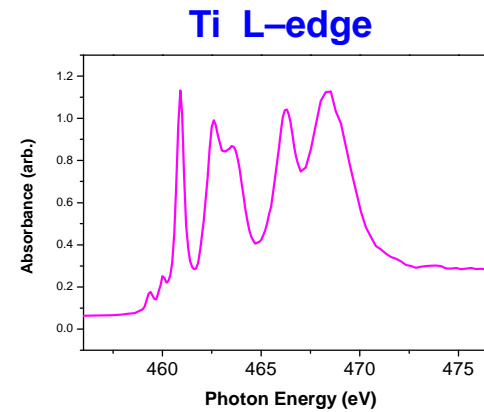
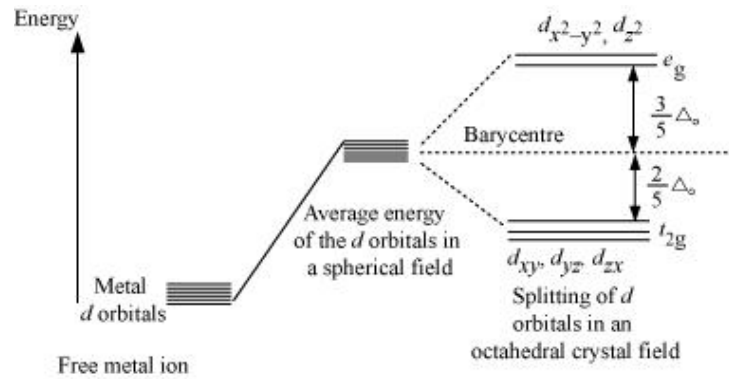


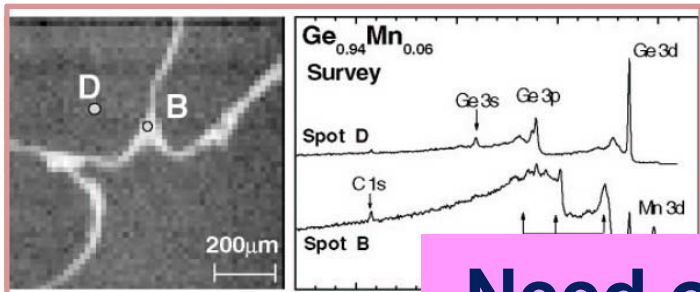


# 1. Practical use of XPS & XAS spectroscopy...

element, crystal structure, chemical states, oxidation state, magnetic moment, electronic structure, ...

W. C. Chueh et al, Sandia Report, 2012.

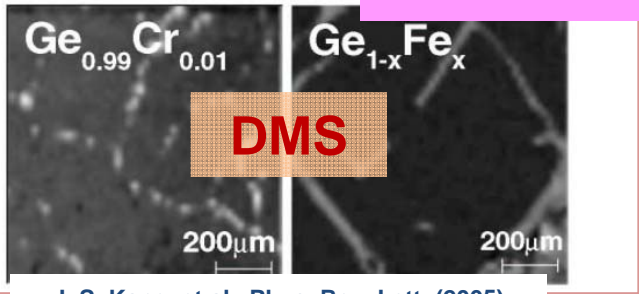




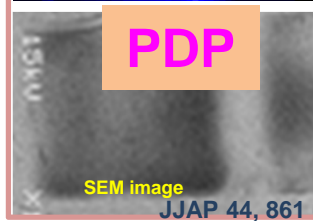
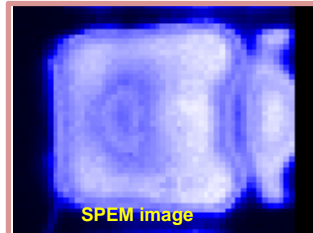
Optoelectronic device

K.J. Kim et al., Adv. Mater. 20, 3589 (2008)  
H. Lim, et al., ChemComm 47, 8608 (2011)

# Need of spectro-microscopy

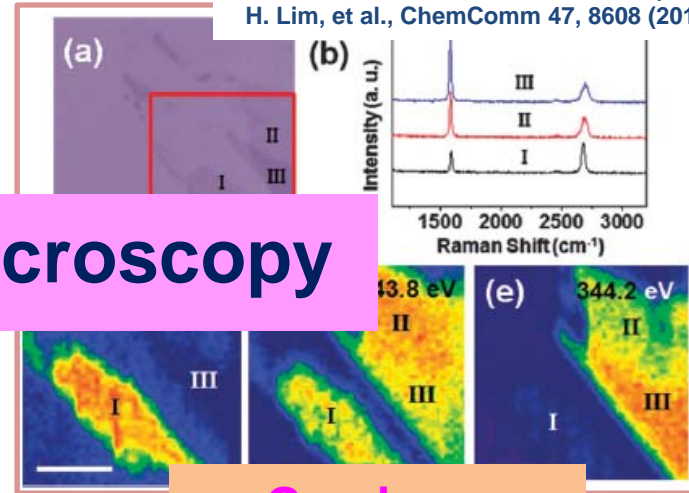


J. S. Kang, et al., Phys. Rev. Lett. (2005)

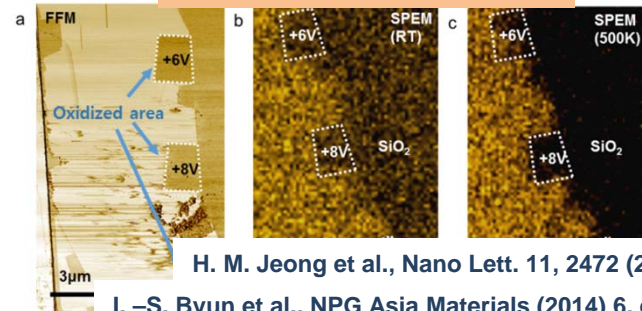


PDP

JJAP 44, 861 (2005)

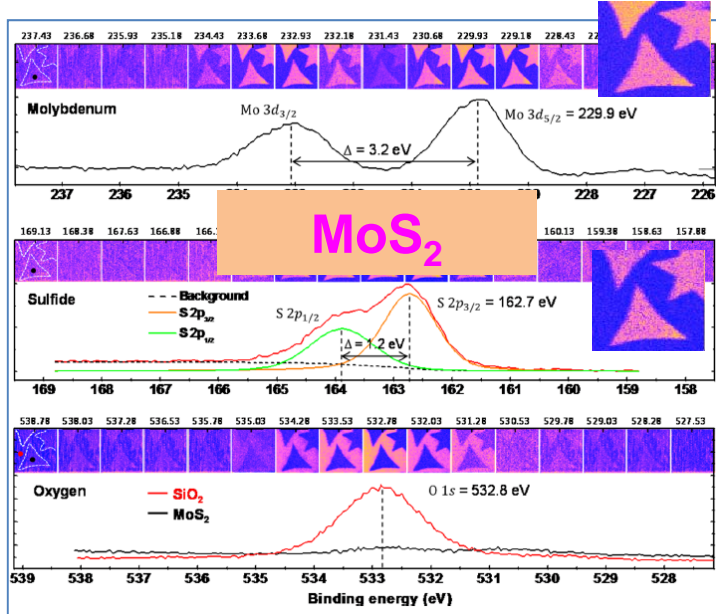


Graphene



H. M. Jeong et al., Nano Lett. 11, 2472 (2011)

I.-S. Byun et al., NPG Asia Materials (2014) 6, e102



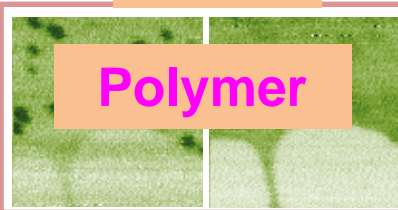
I. Song et al., Angew. Chem. Int. Ed. 53, 1266 (2014)

W. Park et al., ACS Nano 8, 4961 (2014)

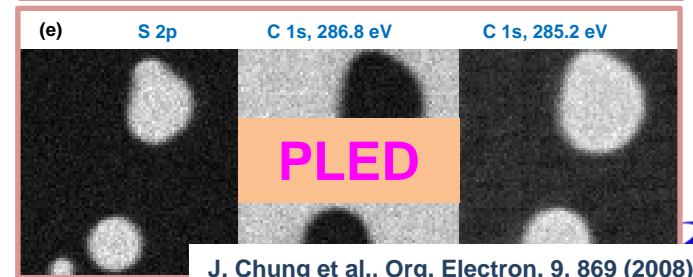
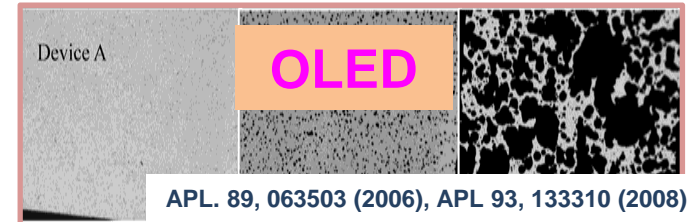
H. J. Shin



Kevlar

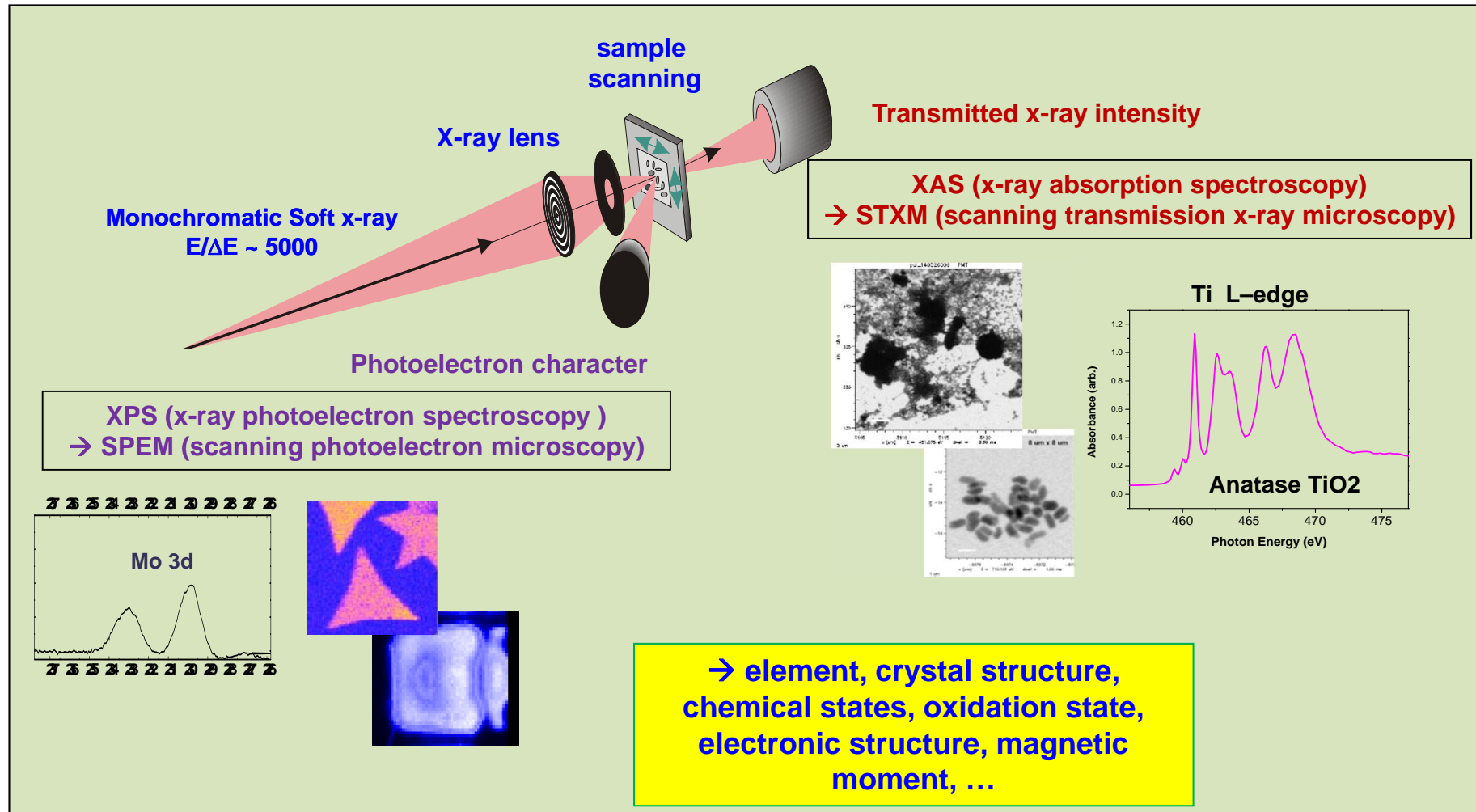


Polymer



PLED

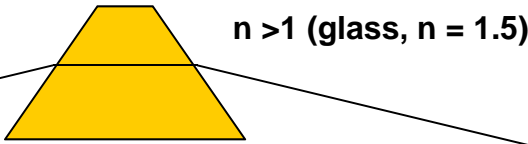
# 1. Spectromicroscopy schematic



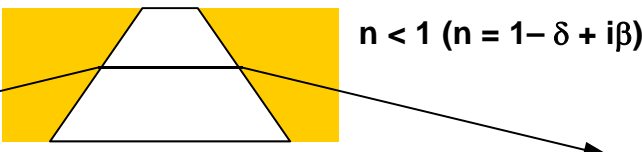
# 1. Lenses for microscopy

Diffraction limit  $\sim 1.22 \lambda$

visible light



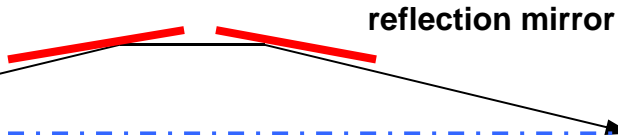
hard x-ray



$\delta$ ; phase shift,  $\beta$ ; absorption  
 $k = n/c, E_0 e^{-i(\omega t - kr)}$

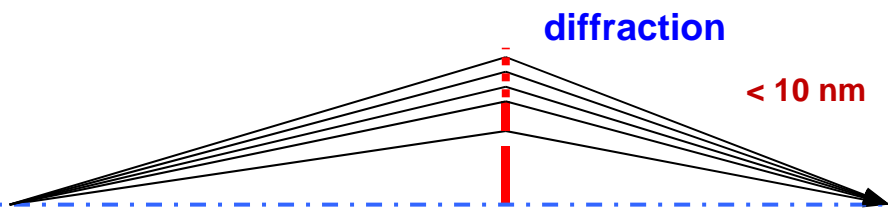
Snigirev, *Nature* 384, 49 (1996)  
 ...

visible, soft & hard x-ray



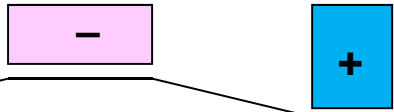
Mimura, *Nat. Phys.* (2010)

visible, soft & hard x-ray



E. Fabrizio, *Nature* 401, 895 (1999)  
 L. Kipp, *Nature* 414, 184 (2001)  
 Y. Wang, *Nature* 424, 50 (2003)  
 W. Chao, *Nature* 435, 1210 (2005)  
 H. C. Kang, *PRL* 96,127401 (2006)

e-beam



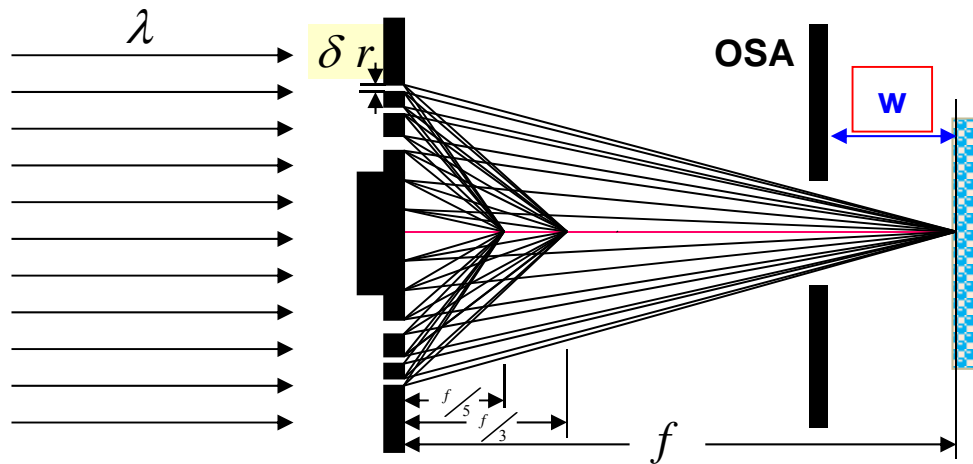
$$\lambda = h / (2mE)^{1/2}$$

$$\lambda [\text{A}] = 12 / [E(\text{eV})]^{1/2}$$



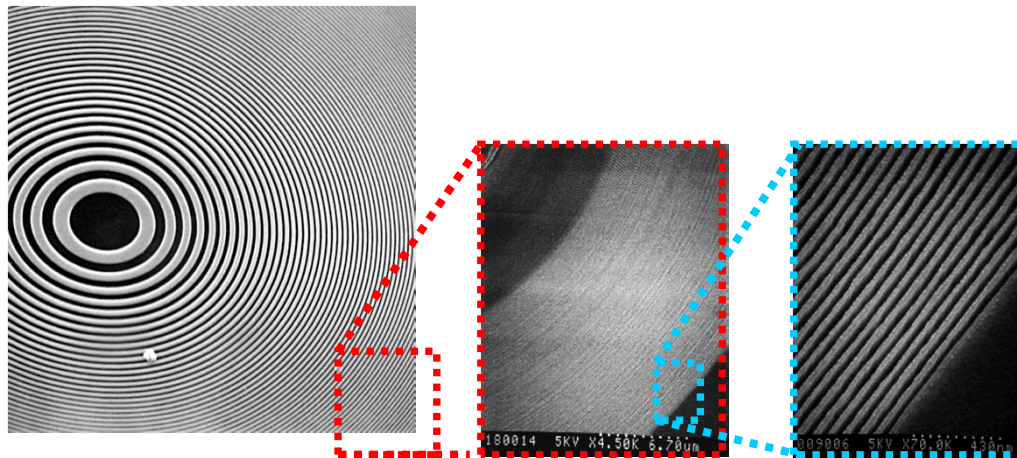
# 1. zone plate as a focusing lens (< 10 nm)

Focal length: depends on E.  
Order sorting aperture (OSA) is needed: short working distance (w).

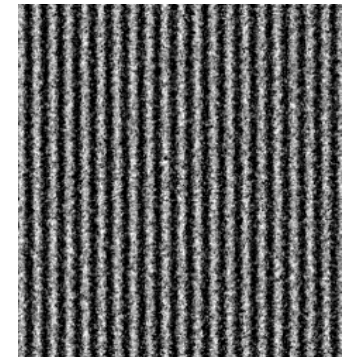


$$f = \frac{4N(\delta r)^2}{\lambda}$$

$$\text{S.R.} = 1.22 \times \delta r$$

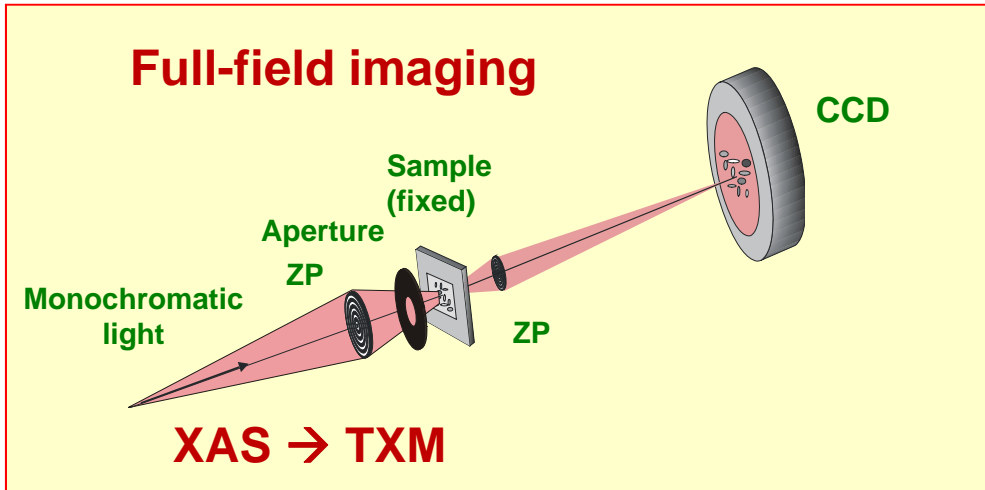


Soft x-ray image of  
15 nm Cr/Si lines & spaces  
W. Chao, *Nature* 435, 1210 (2005)

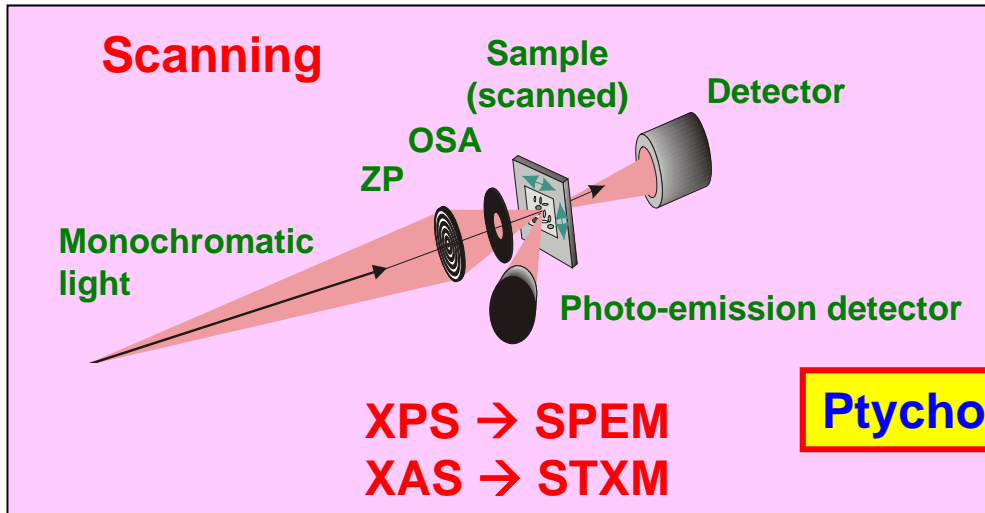
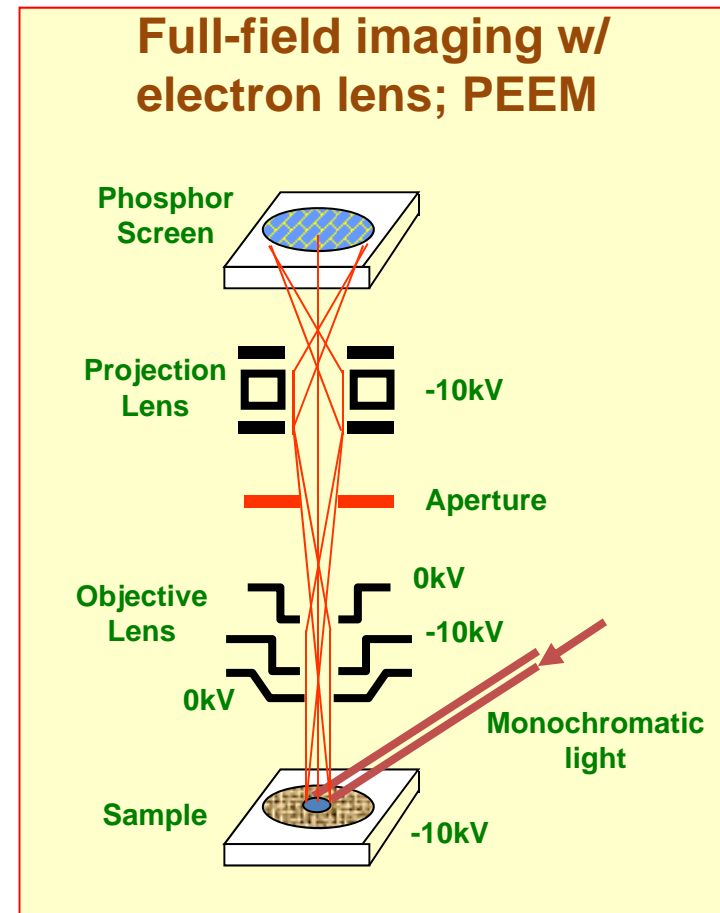




# 2. Soft X-ray (spectro)-Microscopes



schematic: [www.elettra.trieste.it](http://www.elettra.trieste.it)



**Ptychography**

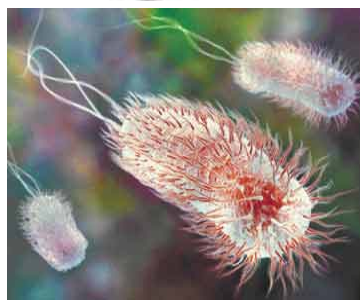
**Lensless imaging: CDI**

# 2.1 TXM (transmission x-ray microscopy)

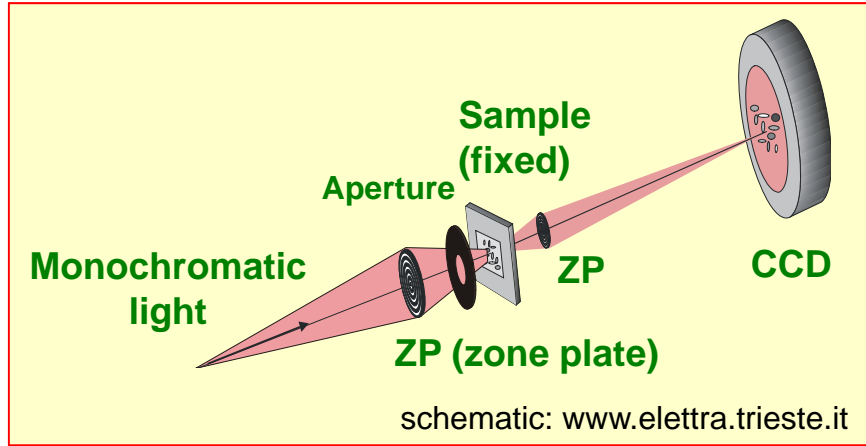
Nondestructive, element specific, ...  
 Spatial resolution: 20 ~ 50 nm.  
 Advantage in bio sample investigation: - no additive for contrast, - internal structure visualization, - 3D  
 CT data acquisition time is shorter than other tools

- easy alignment and operation
- sample can be in atmospheric pressure ...

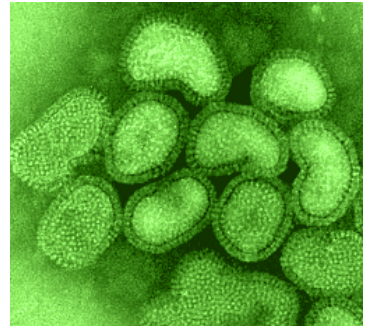
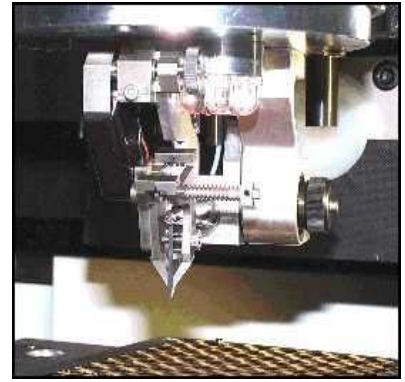
Light microscope



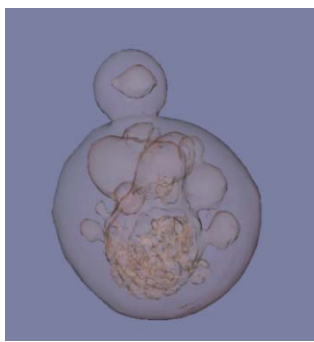
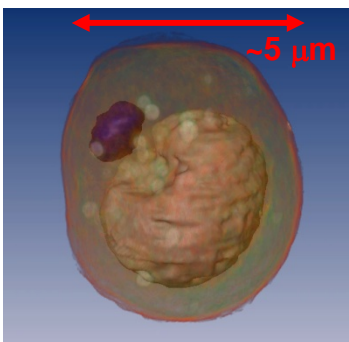
200 ~ 1000 nm



Electron microscope



< 1 nm



Larabell & Le Gros, Mol. Biol. Cell, 15, 957 (2004)

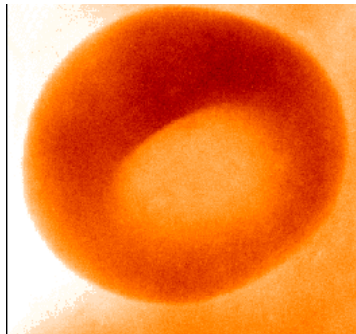


## 2.1 TXM application examples:

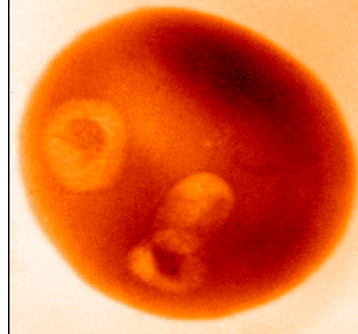
### Malaria infected blood cells

Life cycle of malaria in human red blood cells.

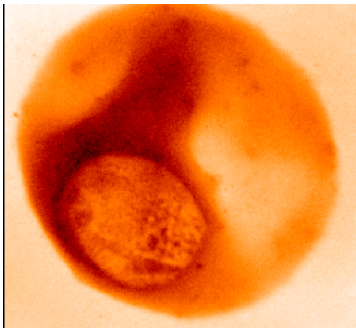
uninfected blood cell



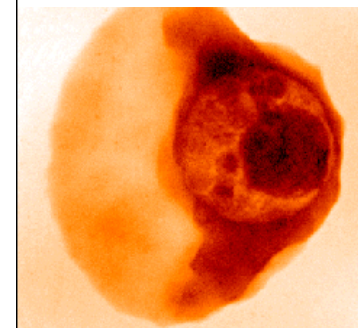
a newly infected cell



a 12- hour old parasite



Leupeptin treated cell

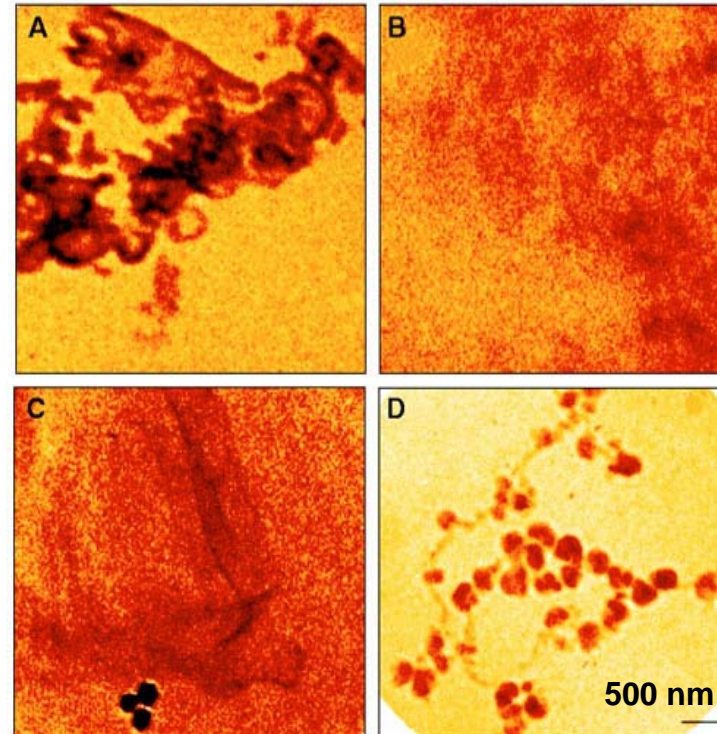


Tubular structure protruding from the parasite into the red blood cell cytosol seen with a young parasite.

Image size = 7  $\mu\text{m}$  x 7  $\mu\text{m}$

<http://www-cxro.lbl.gov>

### Humic substances: how would they dissolve in the soil?



Effect of solution pH.

- A) Globular and ring-like structures in acidic solution.
- B) Uniform, small aggregates in alkaline solution

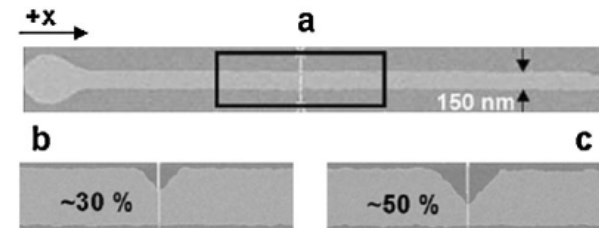
Effect of cation presence.

- C) Thread-like structures with divalent cations.
- D) Globular and thread-like structures with trivalent cations.

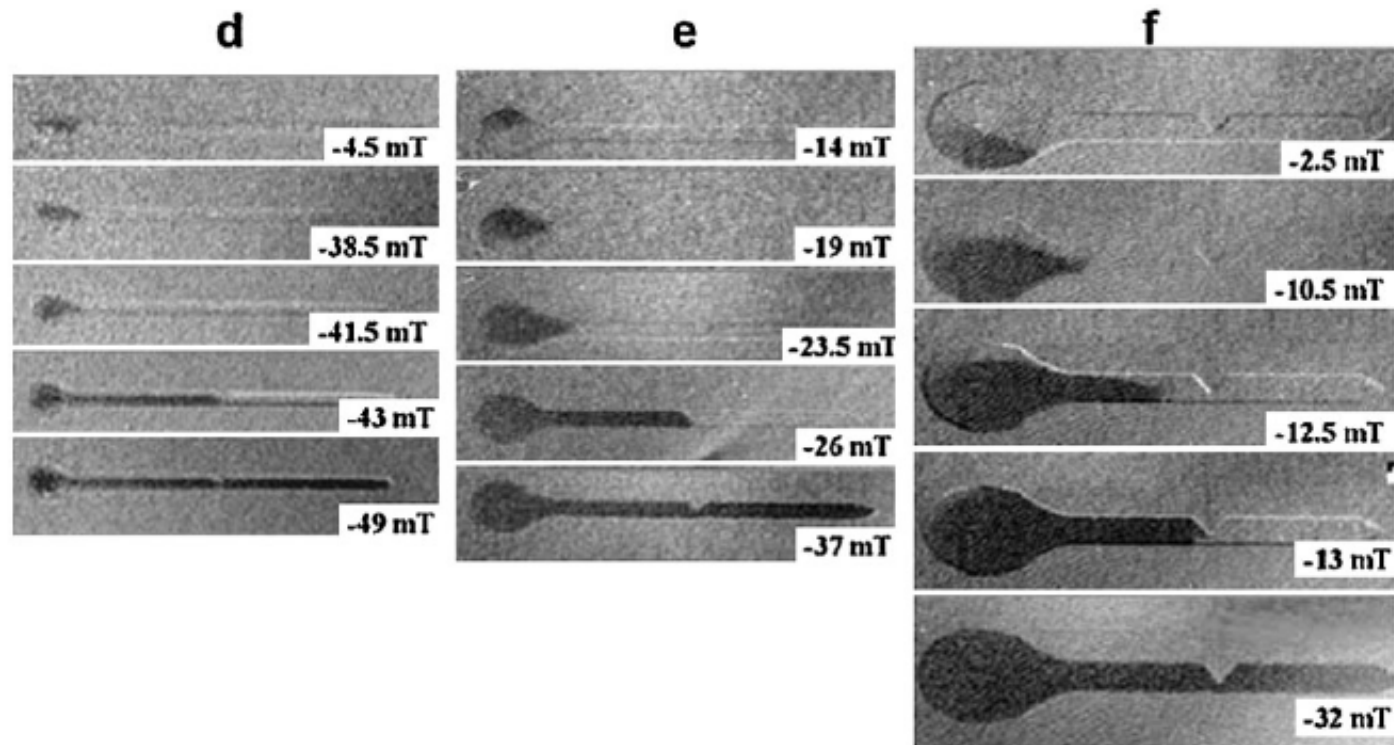
S. Myneni, Science 286, 1335 (1999).

### Direct observation of stochastic domain-wall depinning in magnetic nanowires

(a) Typical SEM image of a 50 nm thick nanowire with a width of 150 nm together with enlarged notch patterns with notch depths of about 30% (b) and about 50% (c) of the wire width.



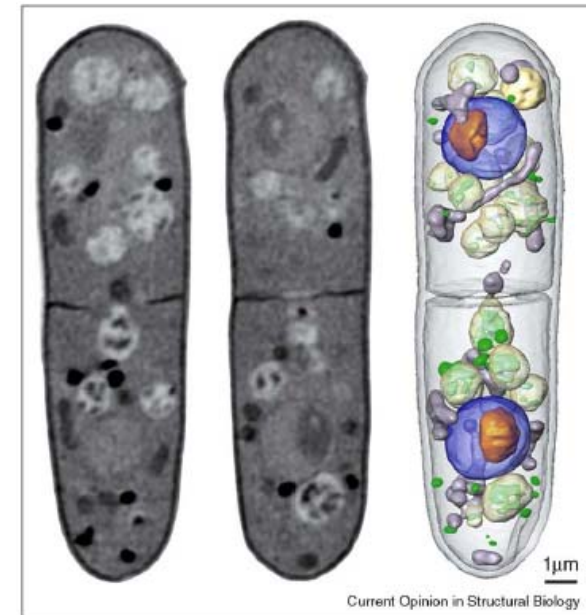
Three representative image sequences of magnetic domain-wall evolution along the hysteresis cycle for wire widths of  $w \approx 150$  nm (d), 250 nm (e), and 450 nm (f). The magnetic field of the DW evolution pattern is indicated on the lower right.



## 2.1 TXM application examples: 3D

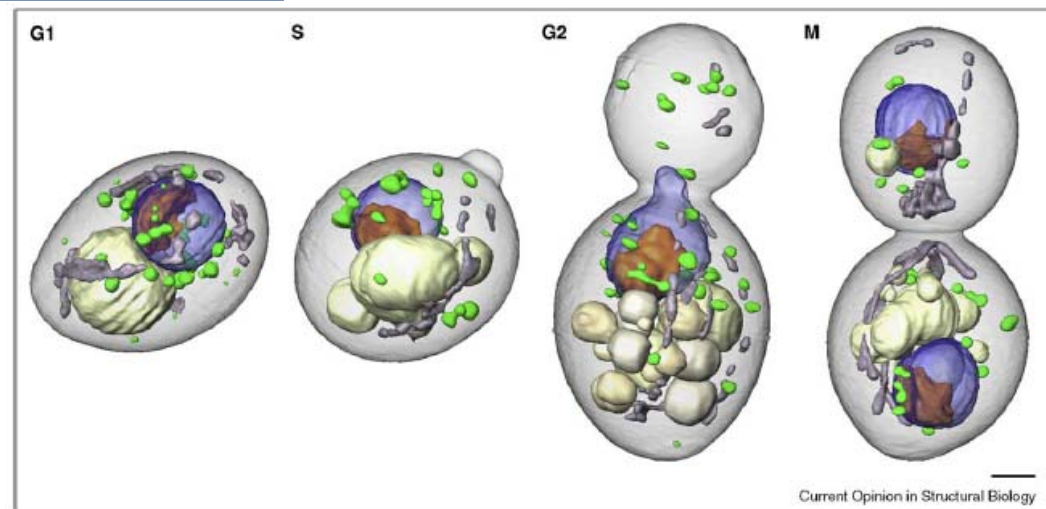
### Rapidly frozen fission yeast

Soft X-ray tomography of a rapidly frozen fission yeast, *Schizosaccharomyces pombe*. The data for the complete three-dimensional reconstruction were composed of 90 images, collected through a total of 180° of rotation. Two computer-generated sections through the tomographic reconstruction are shown along with an image showing select organelles that have been segmented and color-coded according to type. Key: nucleus, blue; nucleolus, orange; mitochondria, gray; vacuoles, white; lipid-rich vesicles, green. (X-ray tomographic reconstructions are an update of the work outlined in [27,28\*\*].)



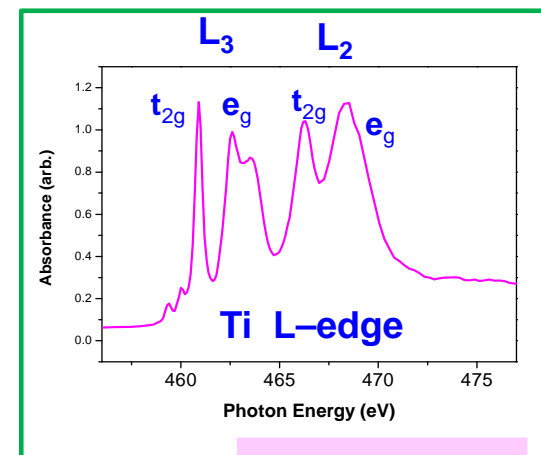
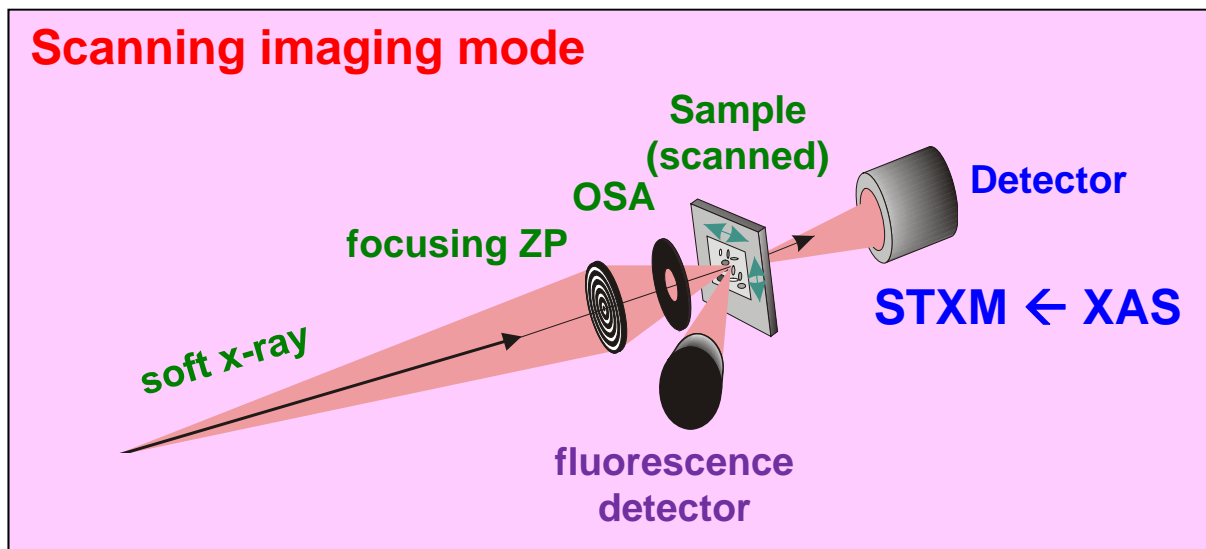
### Rapidly frozen *Saccharomyces cerevisiae* cells

Soft x-ray tomography of rapidly frozen *Saccharomyces cerevisiae* cells imaged at each phase of cell cycle – G1, S, M, and G2. Organelles are color-coded as follows: blue, nucleus; orange, nucleolus; gray, mitochondria; ivory, vacuoles; green, lipid bodies. Scale bar, 2 μm.

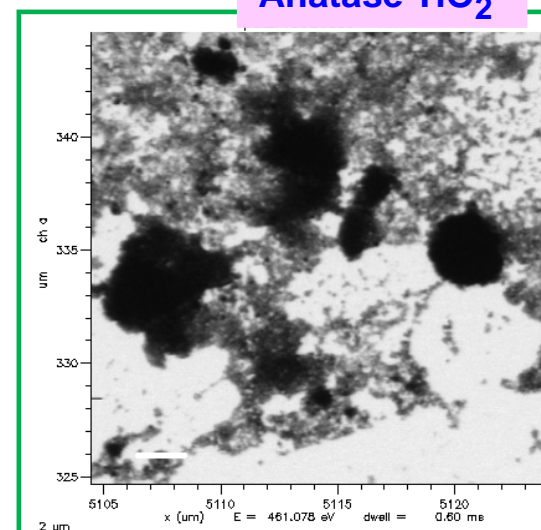




## 2.2 STXM (scanning transmission x-ray microscopy)

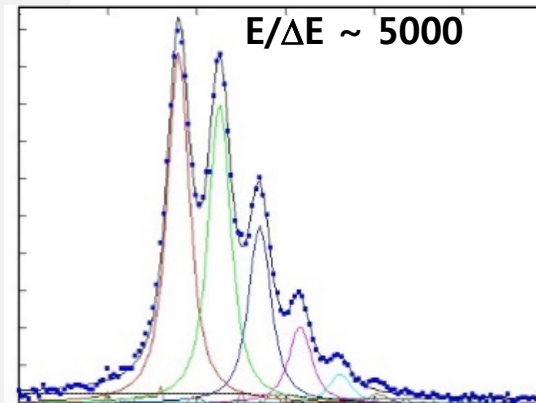
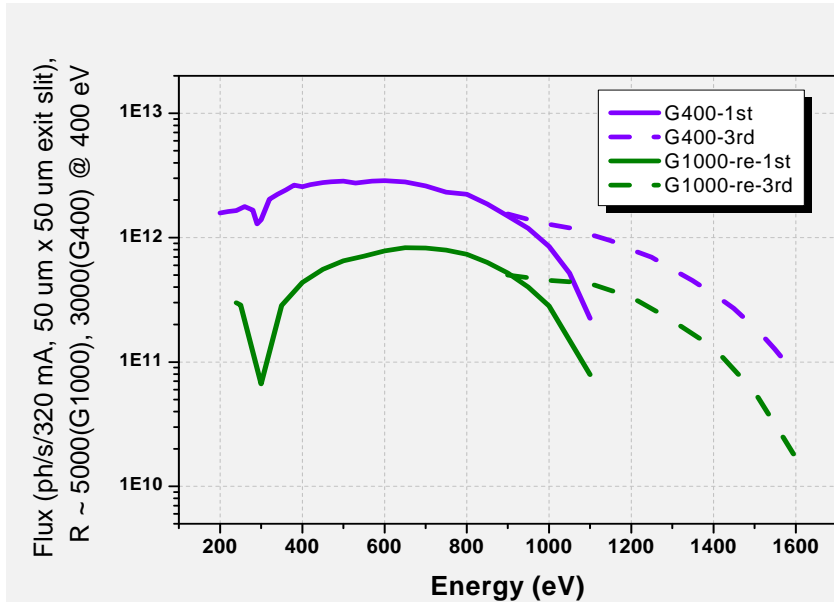
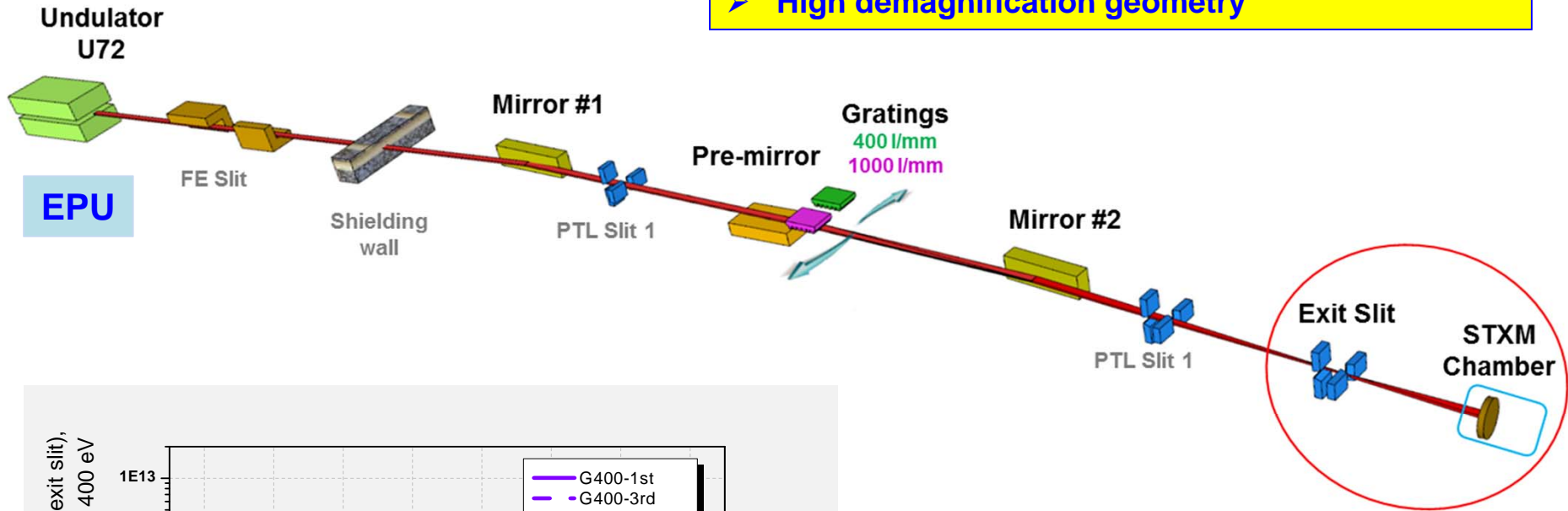


Anatase TiO<sub>2</sub>



## 2.2 STXM requirement: beamline specification

- EPU beamline is better
- Enough flux in wide energy range: 100 – 2000 eV
- Energy resolving power:  $E/\Delta E > 2,000$
- Small virtual source size
- High demagnification geometry



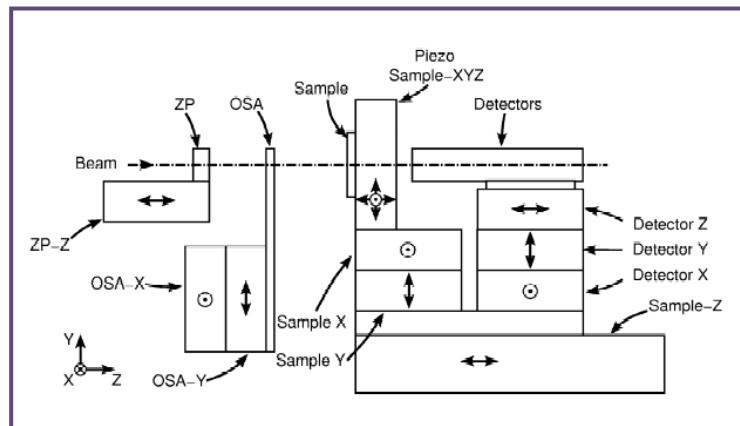
STXM

## 2.2 STXM requirement; apparatus

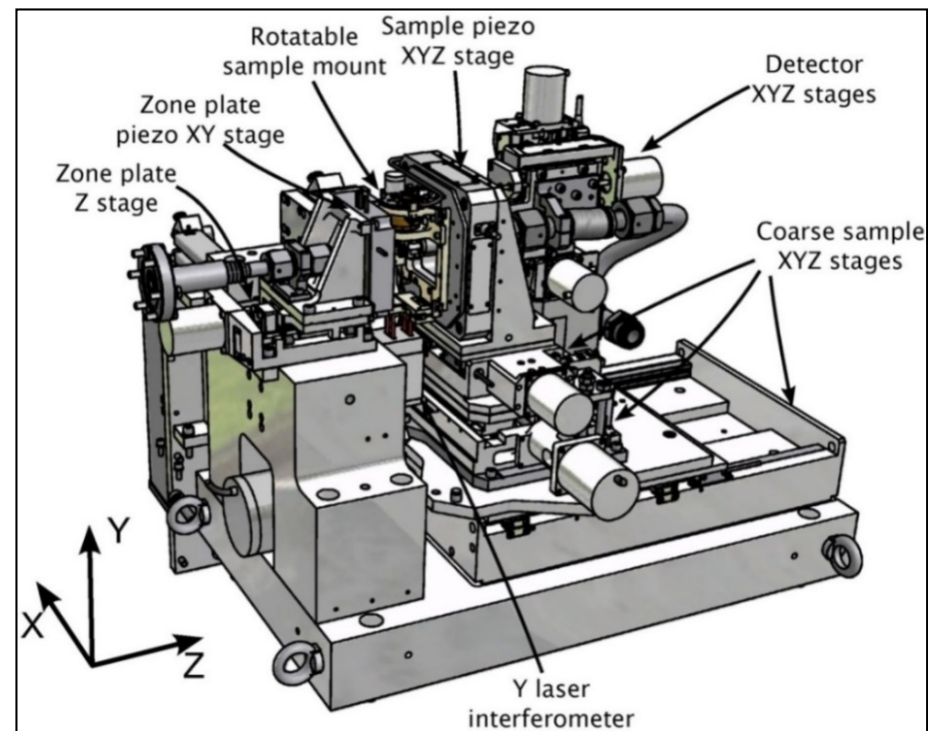
- Accurate and fast scanning stages
- Accurate and fast position monitoring
- Accurate and fast feedback system (Laser interferometry)
- Fast detector

$$f = D \Delta r / \lambda$$

**\*\* OSA**

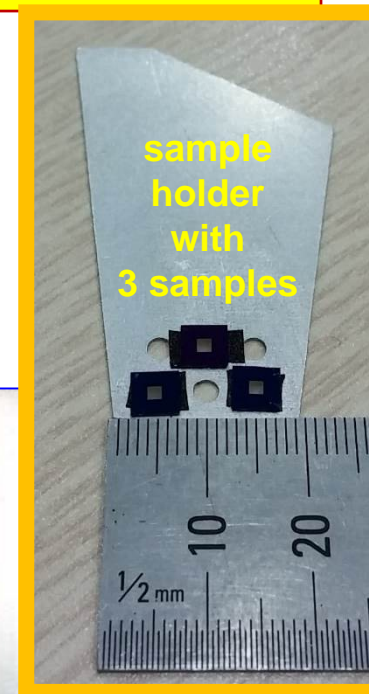
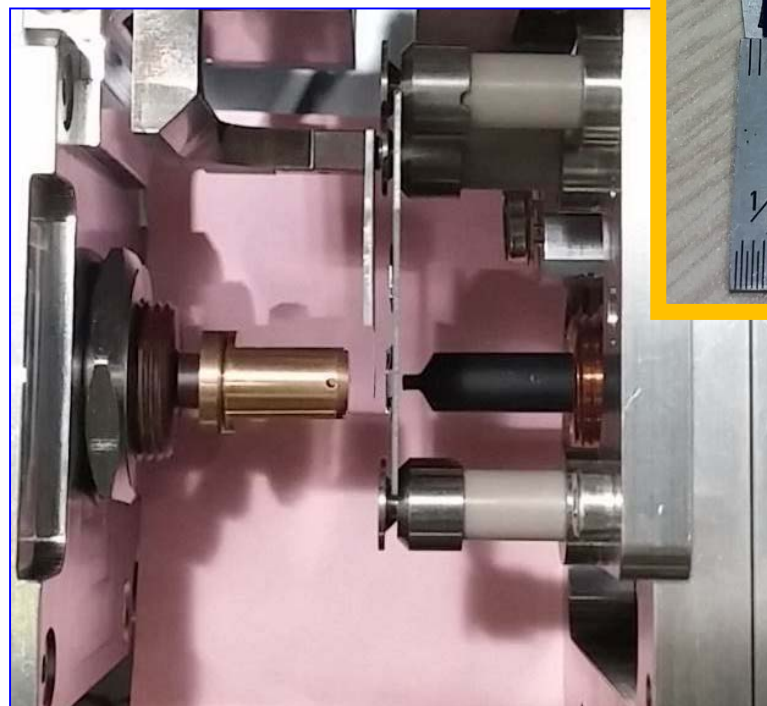
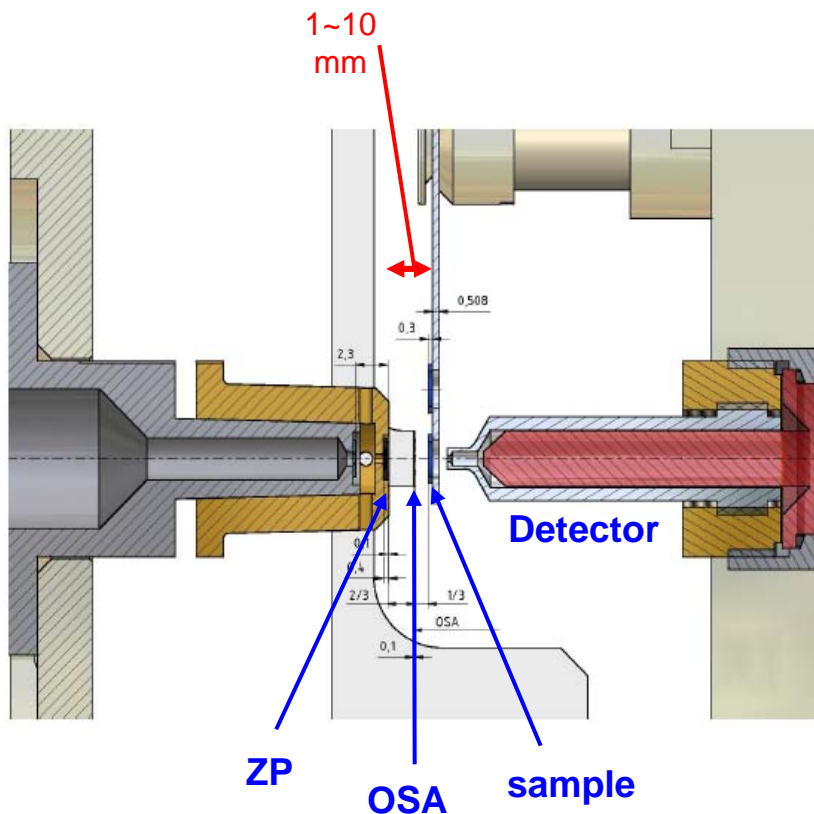


<http://www-als.lbl.gov>  
& Bruker Ltd.



## 2.2 STXM in real shape

- easier to find ROI within sample
- easier to find the best focus
- easier and faster sample loading
- detector is efficient for fast counting...



## 2.2 STXM: single image acquisition

**Image Scan - Estimated Time: 173s**

Image Scan: 250x250 pts, 541.292 eV, 2 ms Dwell, PMT

Scan Type: Image (Line - unidirectional)

Stage: Automatic

Dichroism:  Map Pol EPU Offset,  2 Pol

Defocus: Focus (nm) 31, Auto Defocus

Accel. Dist. (μm) 0.12, Point Delay (ms) 0.08, Auto Parameters

Line Delay (ms) 100, Energy point wait time EPWT (ms) 100, Wait Time Between Regions 200, Additional Wait Time WT=EPWT+dE \* AWT 0

Energies: Current Energy 570.589, Total Points 1, Single Energies  Regions

Energy (eV) 570.589

Continuous Scan with Velocity: 0.012 mm/s Fine Stage

Spatial Regions:  Multiple Regions

Regions	Centre Pos (μm)	Range (μm)	# Points
X	-130.9274	6.0000	250
Y	4941.8943	6.0000	250

Vertical Scan  Auto shutter with points

Scripted Scan  Remote Scan Logging

Sample/Scan Annotation (saved in .hdr file)

Sample X: -133.56, Sample Y: 4939.51, Abs, Data, 1

Beamline Control: Energy (eV) 570.589, Reading 525, Destination 525, Go S

Entrance Slit 0.000, 0, Go S

Exit Slits: Dispersive (μm) 5.4, 0, Go S; NonDispersive(μm) 5.9, 0.3, Go S

EPU Gap [mm] 49.413, 2.40116e, Go

EPU Polarization 0.00, 0.00, 0, Go

EPU Following  On, EPU Harmonic 1st

BL Feedback  On, EPU Offse -0.330, -0.33 set

Exit Slit Curr 0.000, 0.000, Fbk Offse 0.000, set

Slit Curr 0.000, 0.000

Current Scan Status: NEXAFS Line Scan, Image 1 of 0, Time to finish 5m 57s, Region 1 of 1, Elapsed Time: 10m 14s, Line 137 of 137, Next Scan #: 22, Point 100 of 100

Scan Controls: START, Sample Scan, Abort, Pause

Microscope Control: Motor, Current, Destination / Jog, Setup, Motors

EPU Gap 49.414, 0, J Stop S All OFF

CoarseZ 938.3, 823.439, Go D Stop S All ON

Focus: Auto S, Reset Interfer, Sample Mover, OSA IN, Zone Plate IN, Sample IN, Stop All

Acquire lock, OSA OUT, Zone Plate OUT, Sample OUT

Microscope Status: SampleX -134.043, SampleY 4938.895, SampleZ 0.000, CoarseX -122.50, CoarseY 4896.33, CoarseZ 938.3, ZonePlate -1924.85, OSA-X 0.2, OSA-Y 0.0, DET-X 0.9, DET-Y 2.2, DET-Z 0.4, Taper 0.00, Analog Out -5.000, M1\_Pitch -1.2850, M2\_Pitch -6.5970

**Field of view :  
6 μm x 6 μm  
250 x 250 points**



**2.2 STXM stack images: acquisition of point intensity/ line intensity profile/ 2-D image at different photon energy for taking spectral information...**

Image Scan - Estimated Time: 30m 4s

Scan Type: Image (Line - unidirectional)

Stage: Automatic

Dichroism:  Map Pol EPU Offset  2 Pol

Energies

Current Energy: **570.589**

Total Points: **47**

Single Energy Regions: 3  Same dwell

Region	Start Energy (eV)	End Energy (eV)	Range (eV)	# Points	Step (eV)	Dwell Time (ms)
1	525	532	7	8	1	2
2	532	545	13	26	0.5	2
3	545	570	25	13	2	2

Coarse Stage: scl PAL\_140606014 PMT

Velocity: 0.012 mm/s Fine Stage

	Centre Pos (µm)	Range (µm)	# Points	Step (µm)
X	-132.7805	2.5000	100	0.0250
Y	4940.3224	2.5000	100	0.0250

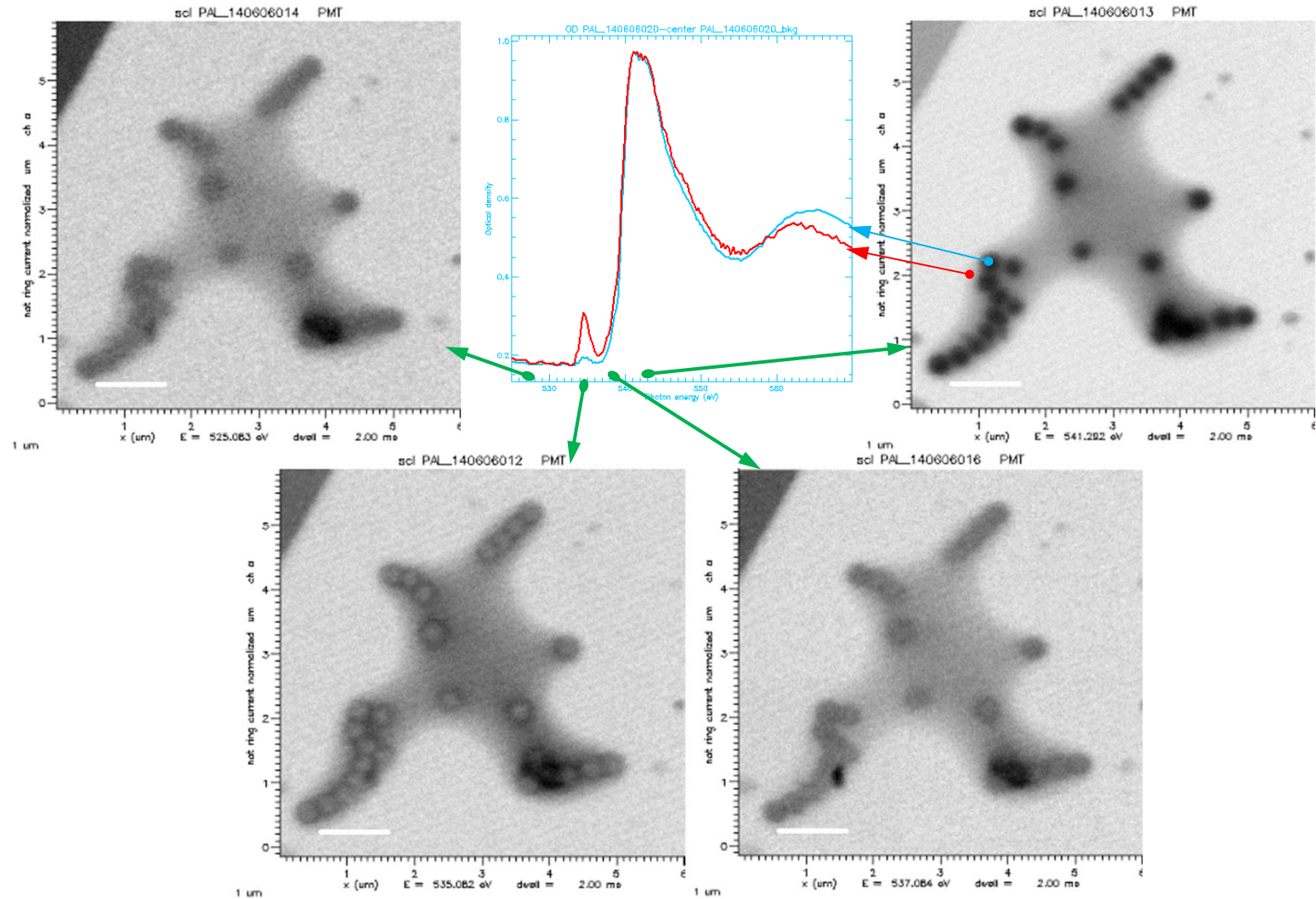
Buttons: **Begin Scan**, Cancel, Save Scan Definition..., Load Scan Definition..., Load Energy Definition..., Recorded Channels..., Check Velocity, Move To First Energy Point, Move to First Position, Reset interferometer, Record Position , Change Magn Field , Energy Move Test

not ring current normalized um

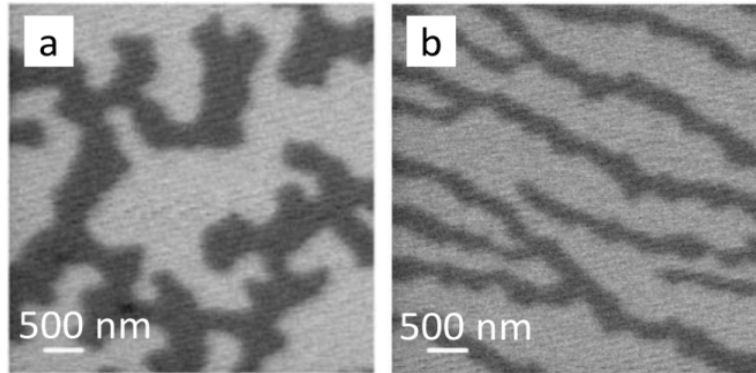
x (um) E = 525.083 eV dwell = 2.00 ms

**2.2 STXM: images at different photon energies & spectra at any point or region of interest.**

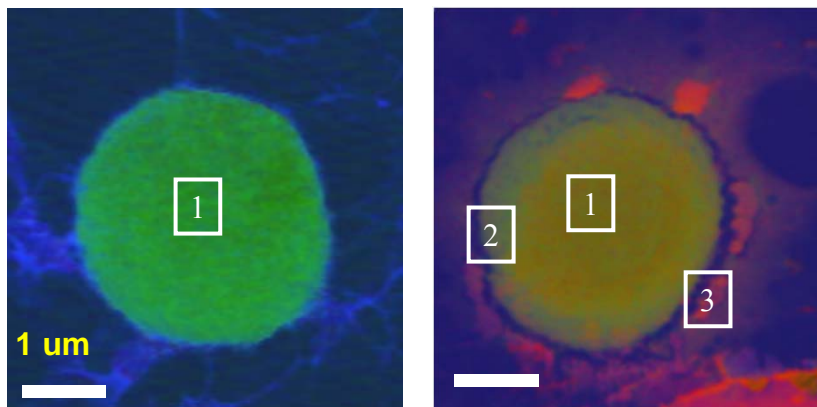
**300 nm silica images at different photon energies & O K-edge spectra from edge & center of silica nano particle**



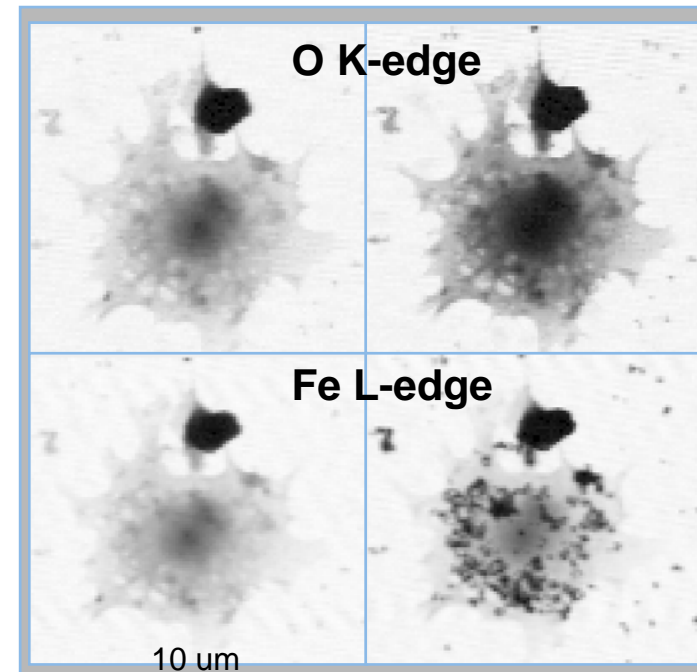
## 2.2 STXM application examples



Courtesy of Prof. D. H. Kim (Chungbuk Univ.)

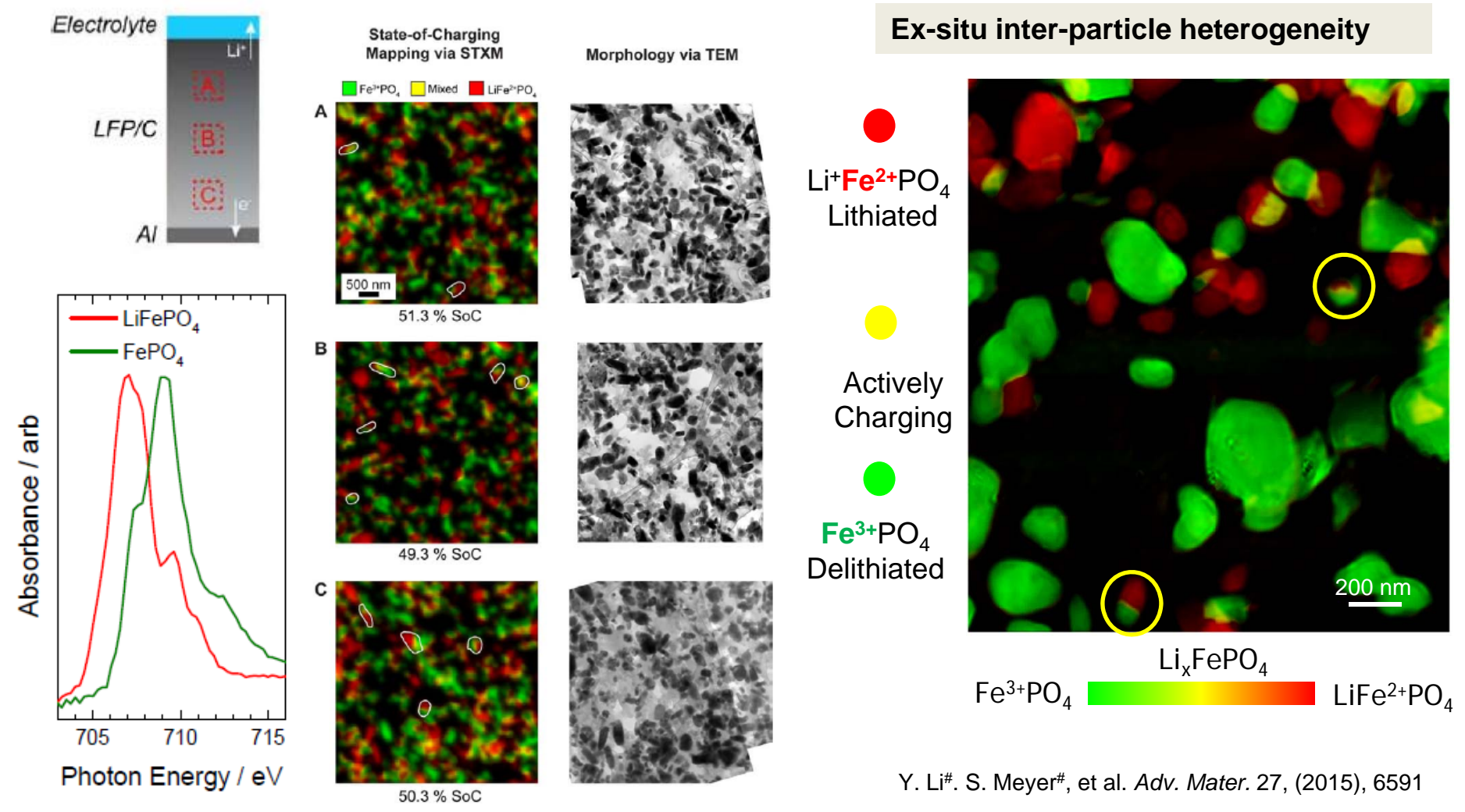


N. Kim, submitted    ■ ROS    ■ Cell    ■ Background



Courtesy of Prof. J. -K. Kim (Cath. Univ.)

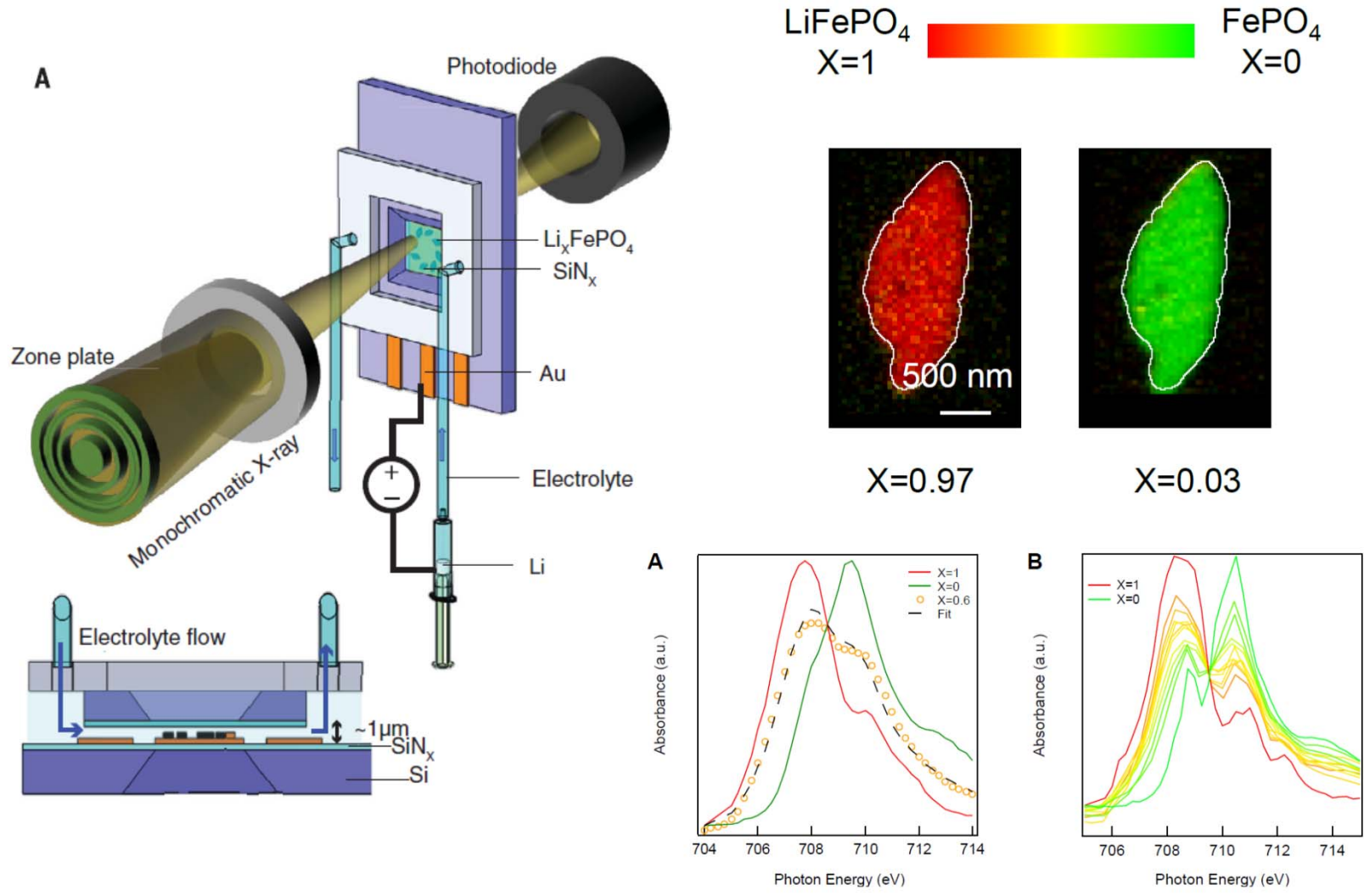
**2.2 STXM application examples : Quasi in-situ study of phase transformation in Lithium-ion batteries**



W. C. Chueh et al. ,  
Sandia Report, 2012.



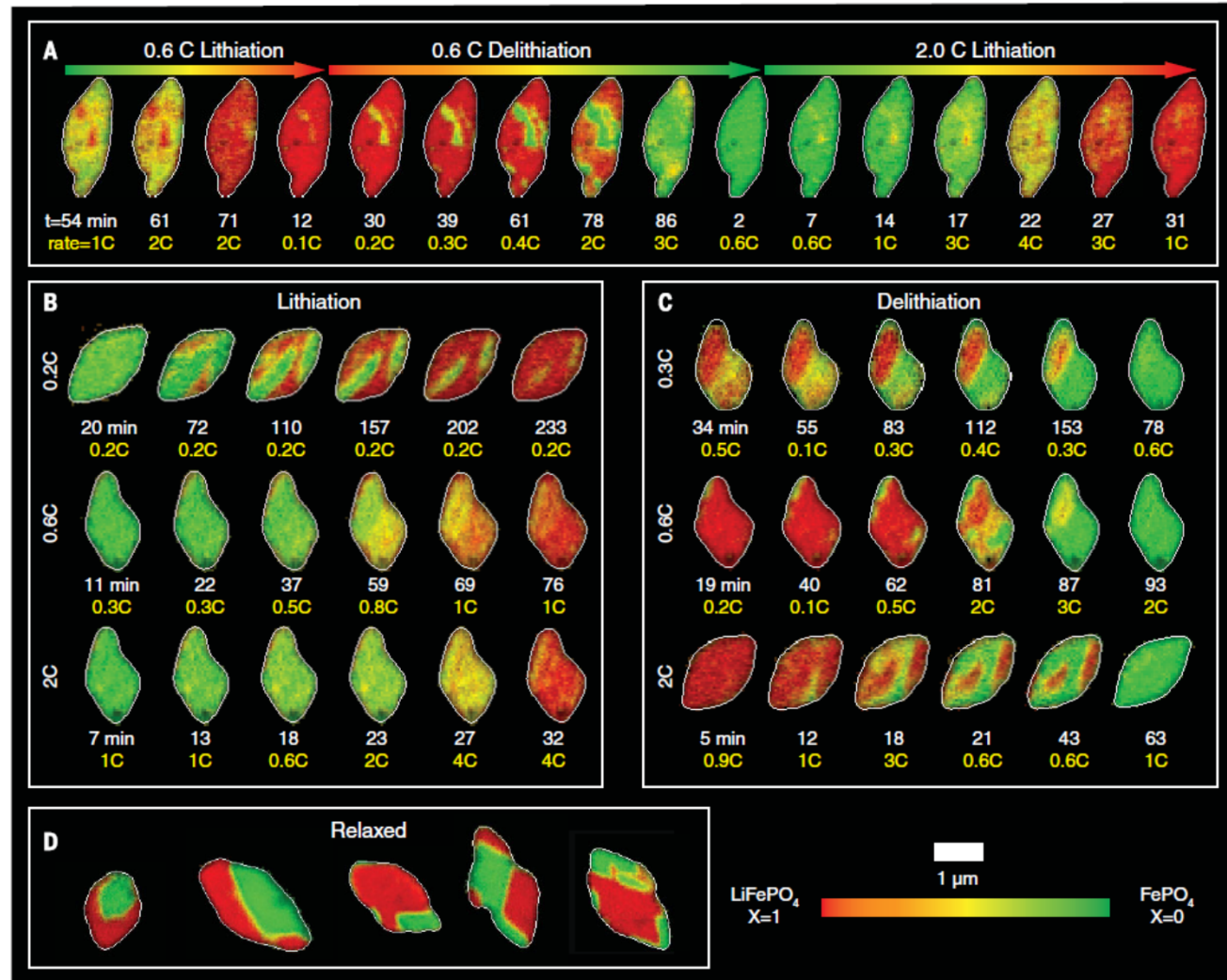
**2.2 STXM application examples: Operando investigation of lithiation and delithiation process of  $\text{Li}_x\text{FePO}_4$ ...**



J. Lim et al., Science, 353, 566 (2016)



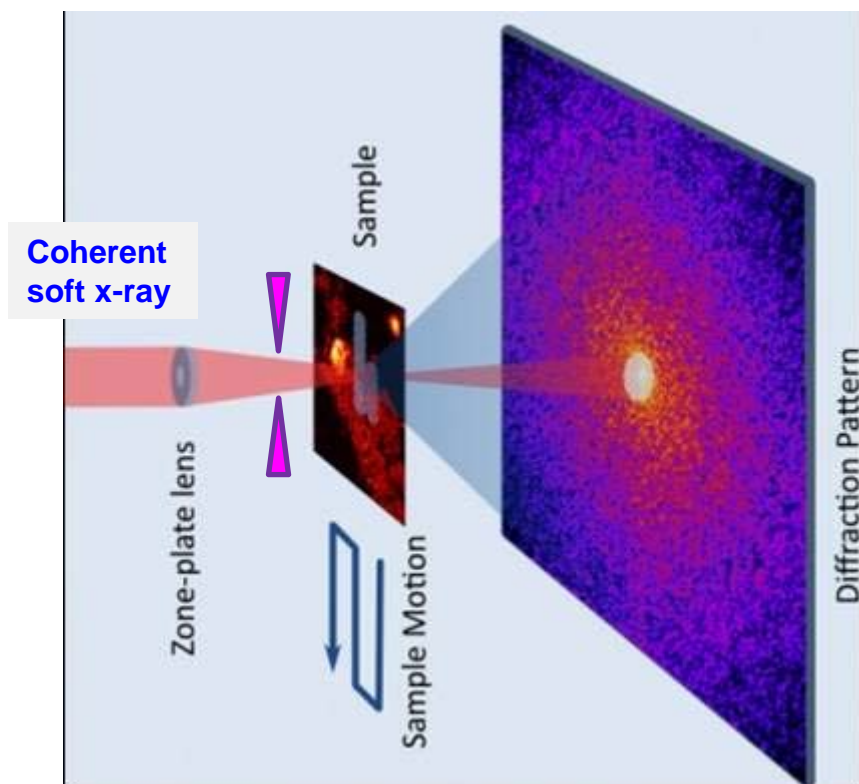
Rate dependent (de) lithiation heterogeneity (0.6 C, 2C)  
 Current density (insertion rate) quantification  
 Exchange current density vs. Li composition  
 Origin of heterogeneity.



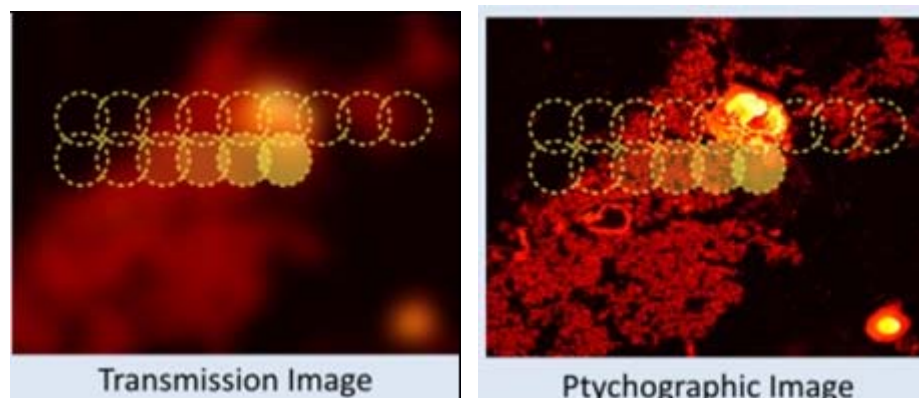
## 2.2 STXM; ptychography mode to improve space resolution...

27

Increase of N.A. :  
~ 2-5 nm space resolution is possible ...



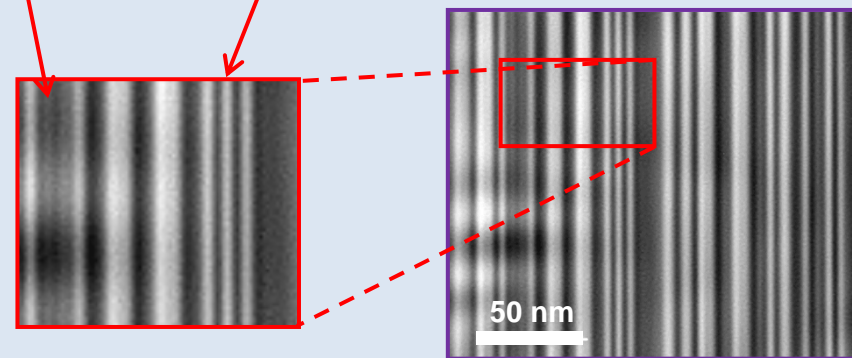
D. Shapiro, T. Tyliszczak et al., Nat. Photonics (2014).  
Nat. Mater. (2014).  
Nano Lett. (2015).



ALS, Berkeley

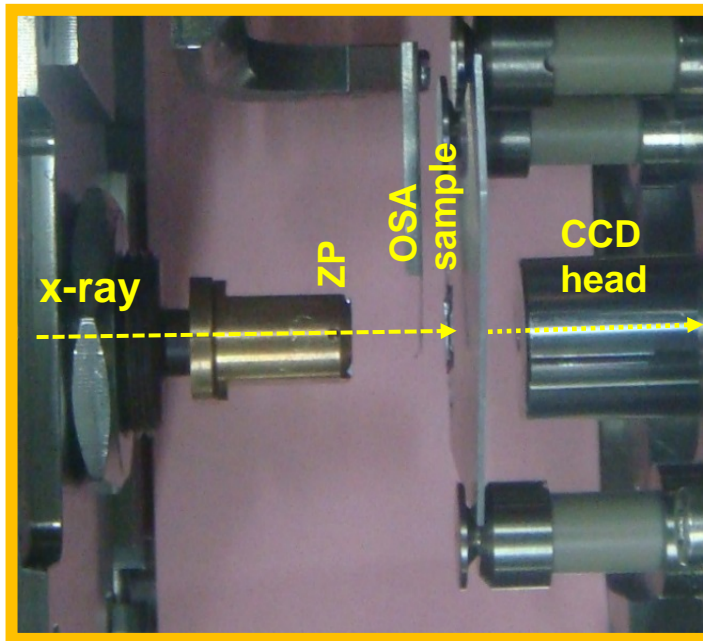
1.5 nm line resolved

3 nm line

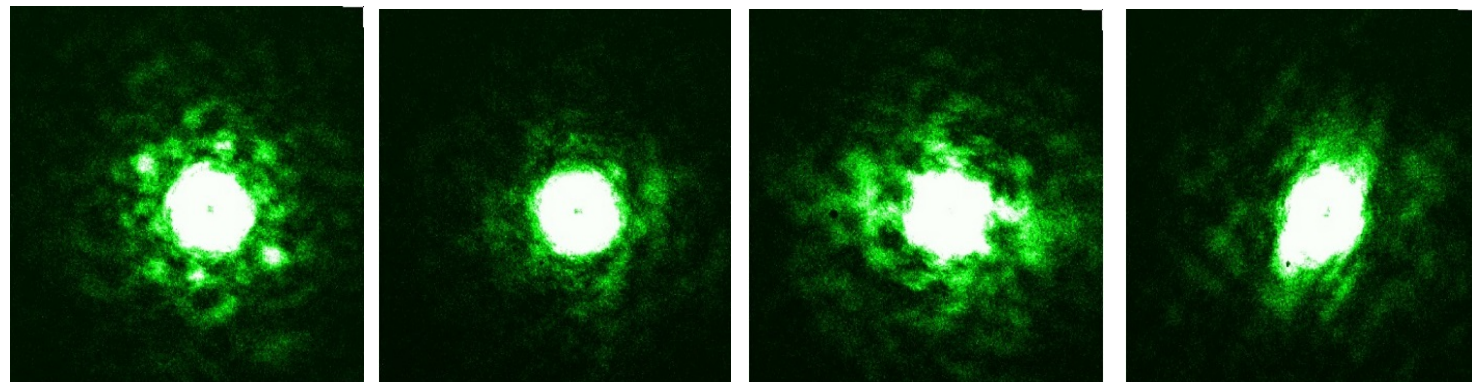
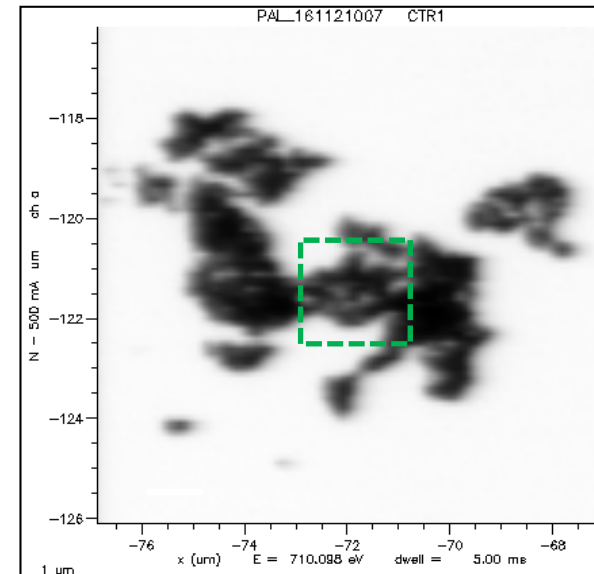


## 2.2 STXM; ptychography setup in practical example

28



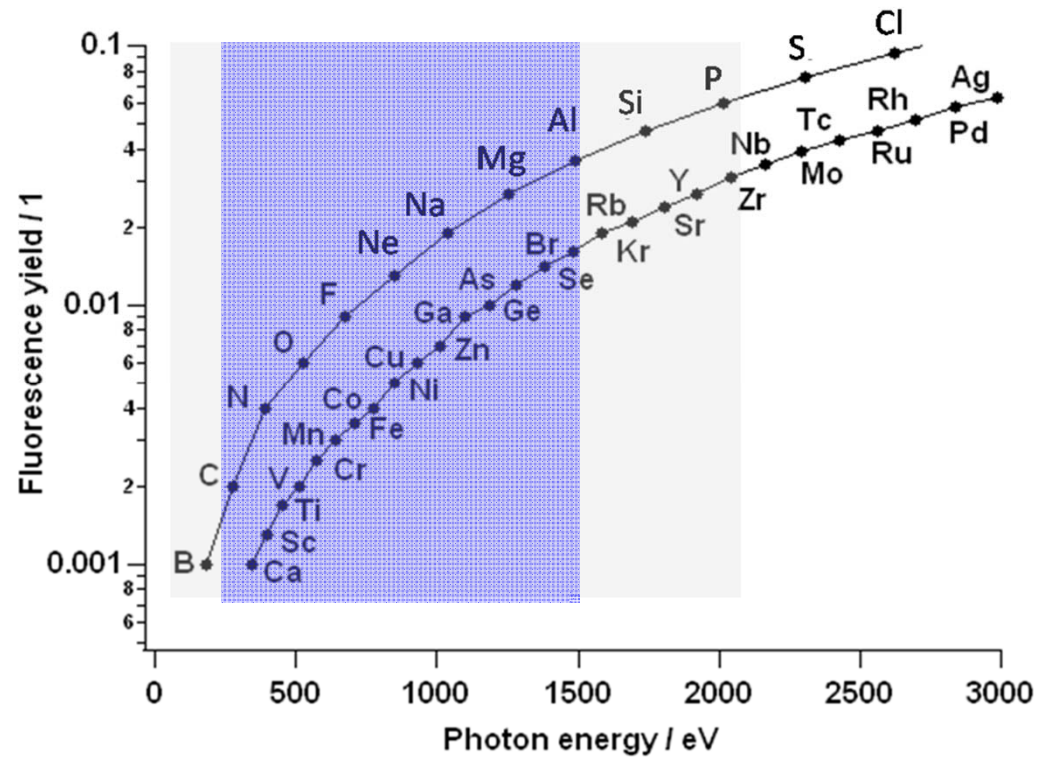
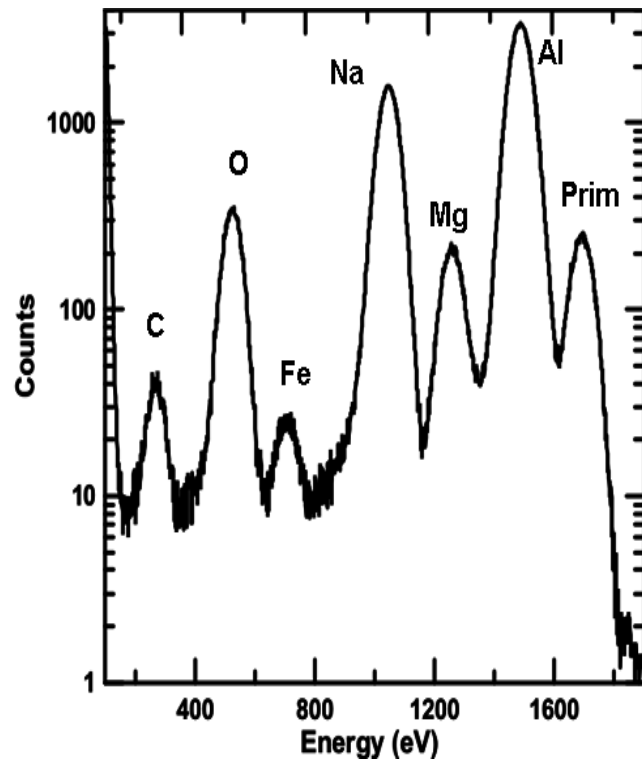
Sample: FeO<sub>x</sub> nanoparticles  
Photon energy: 710 eV  
1  $\mu\text{m}$  x 1  $\mu\text{m}$  (20 x 20 points)  
Beam size at sample =  $\sim$ 70 nm  
Sample to CCD distance =  $\sim$ 80 mm





## 2.2 STXM; fluorescence mode

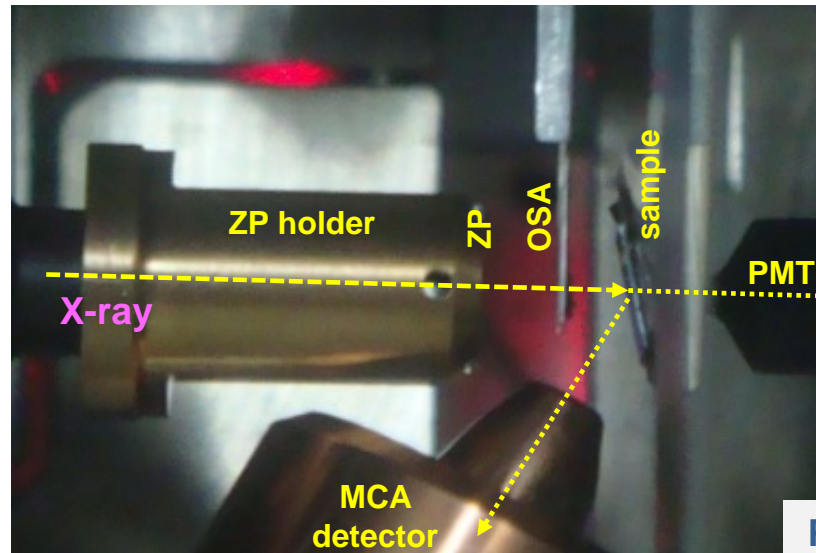
- Fluorescence yield is low for soft X-rays..
- Multi element information in a single scan !
- Low Z elements !
- Can probe thick sample ...



A. Gianoncelli, et al., NIMA 608 (1), 195.

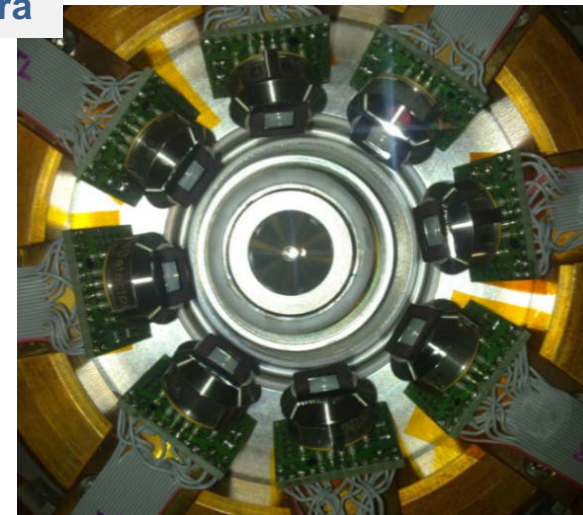
## 2.2 STXM; fluorescence mode in practical application

30



PLS, ALS, CLS

Elettra

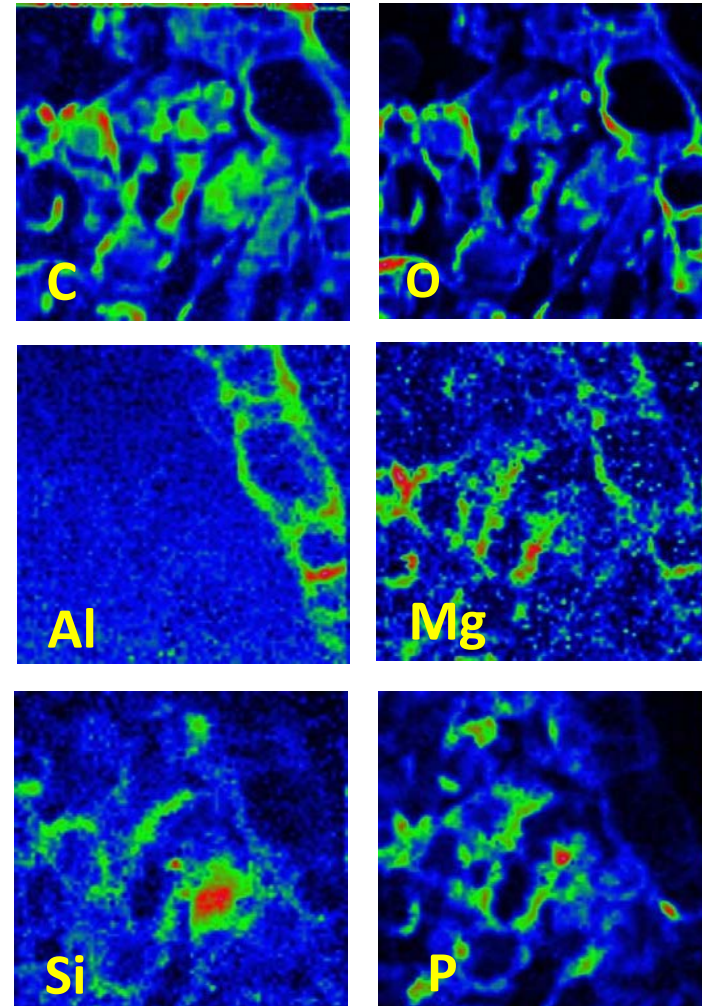
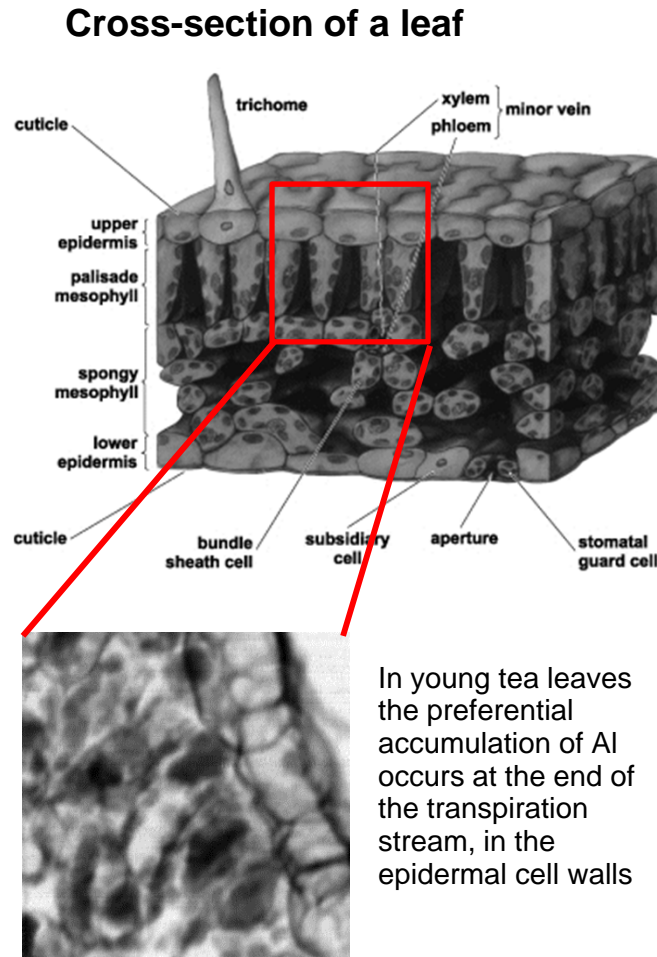


A. Gianoncelli, B. Kaulich, M. Kiskinova, R. Alberti, T. Klatka, A. Longoni, A. de Marco, A. Marcello, Simultaneous Soft X-ray Transmission and Emission Microscopy, *Nucl. Instr. and Meth. A* 608 (1), 195-198

### Al in tea leaves



Functionality and toxicity of Al in tea leaves analyzed on sub-cellular level



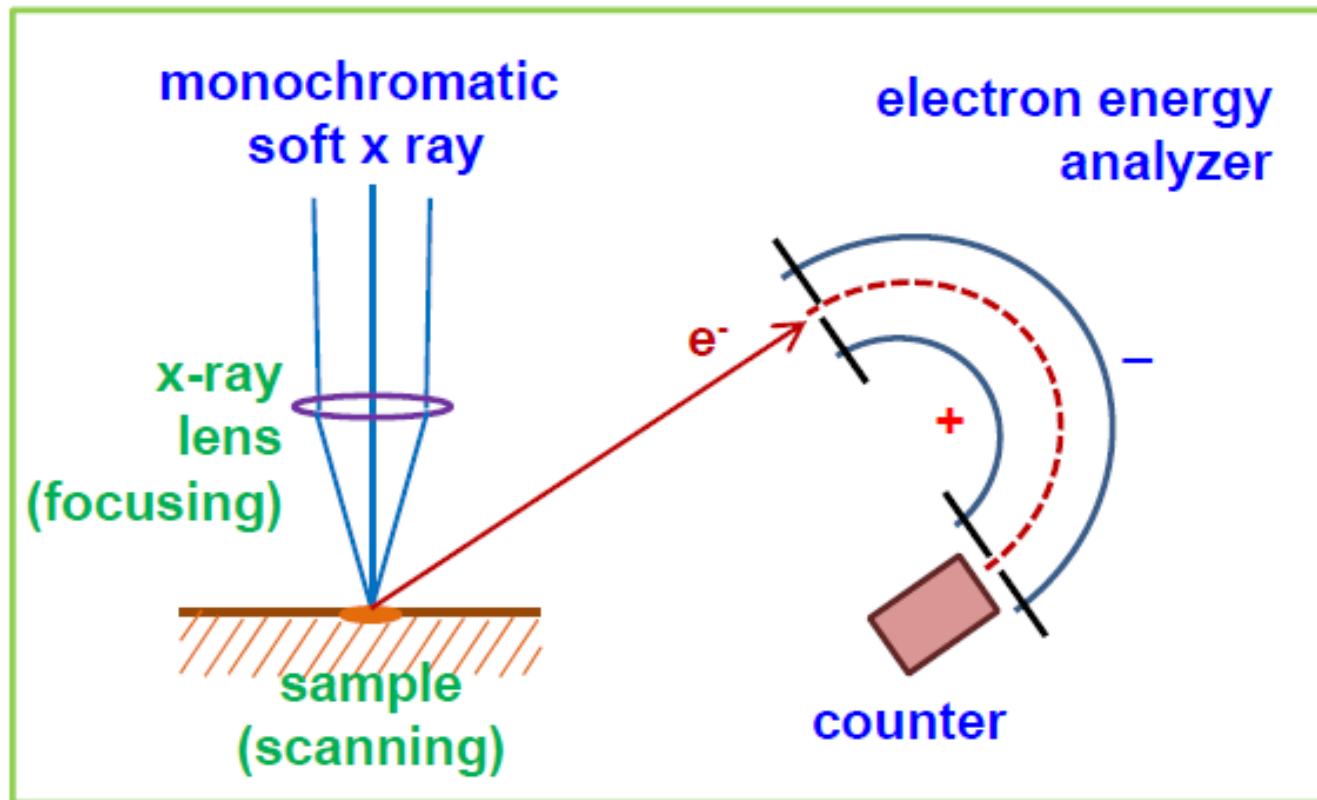
R. Tolra, K. Vogel-Mikus, R. Hajiboland, P. Kump, P. Pongrac, B. Kaulich, A. Gianoncelli, V. Babin, J. Barcelo, M. Regvar, C. Poschenrieder, *Localization of aluminium in tea (Camellia sinensis) leaves using low-energy X-ray fluorescence spectro-microscopy*, J Plant Research 124, 165-172.



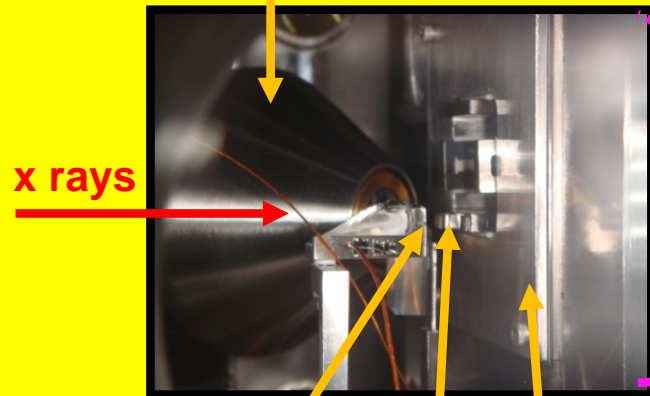
## 2.3 SPEM (Scanning photoelectron microscopy )

### X-ray photoelectron spectroscopy (XPS)

- is a useful probe to investigate chemical states, electronic structure, ...
- is very sensitive to surface.

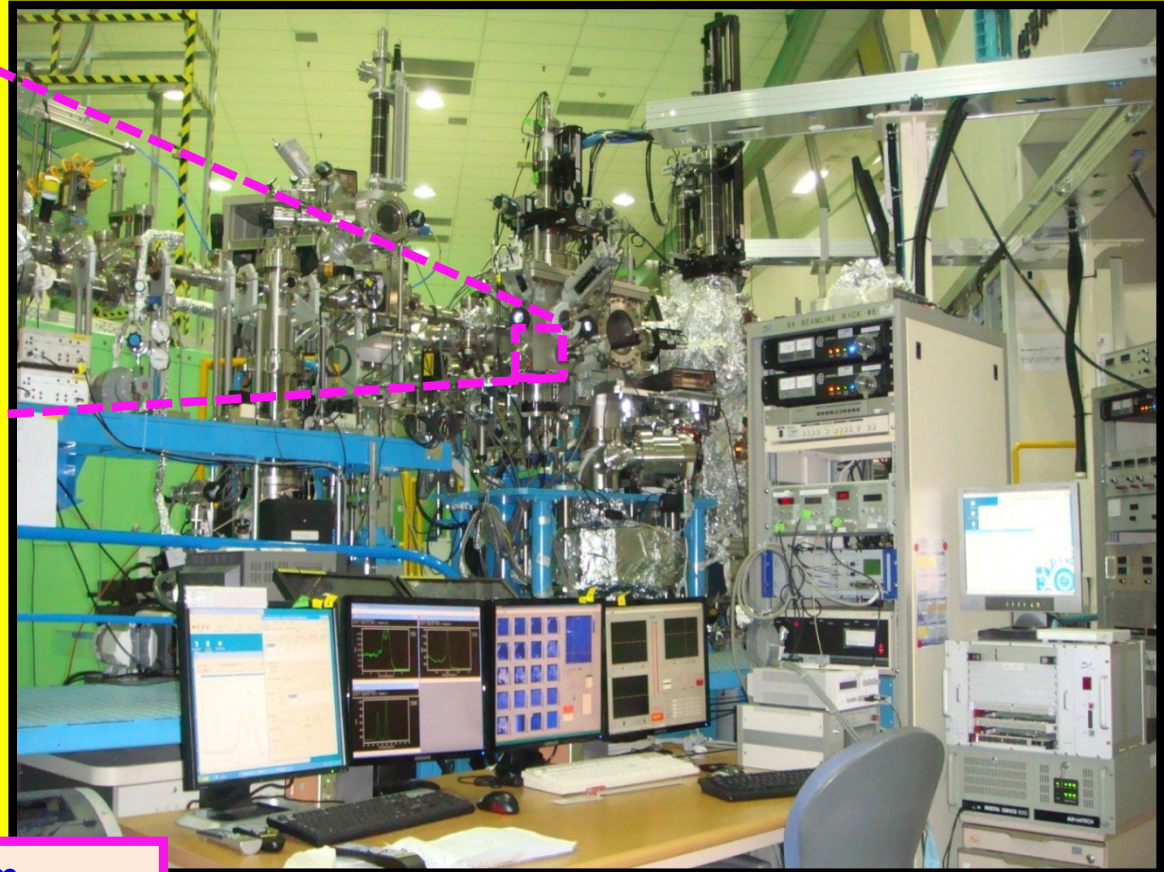


### electron energy analyzer



x-ray lens (ZP)  
+  
OSA

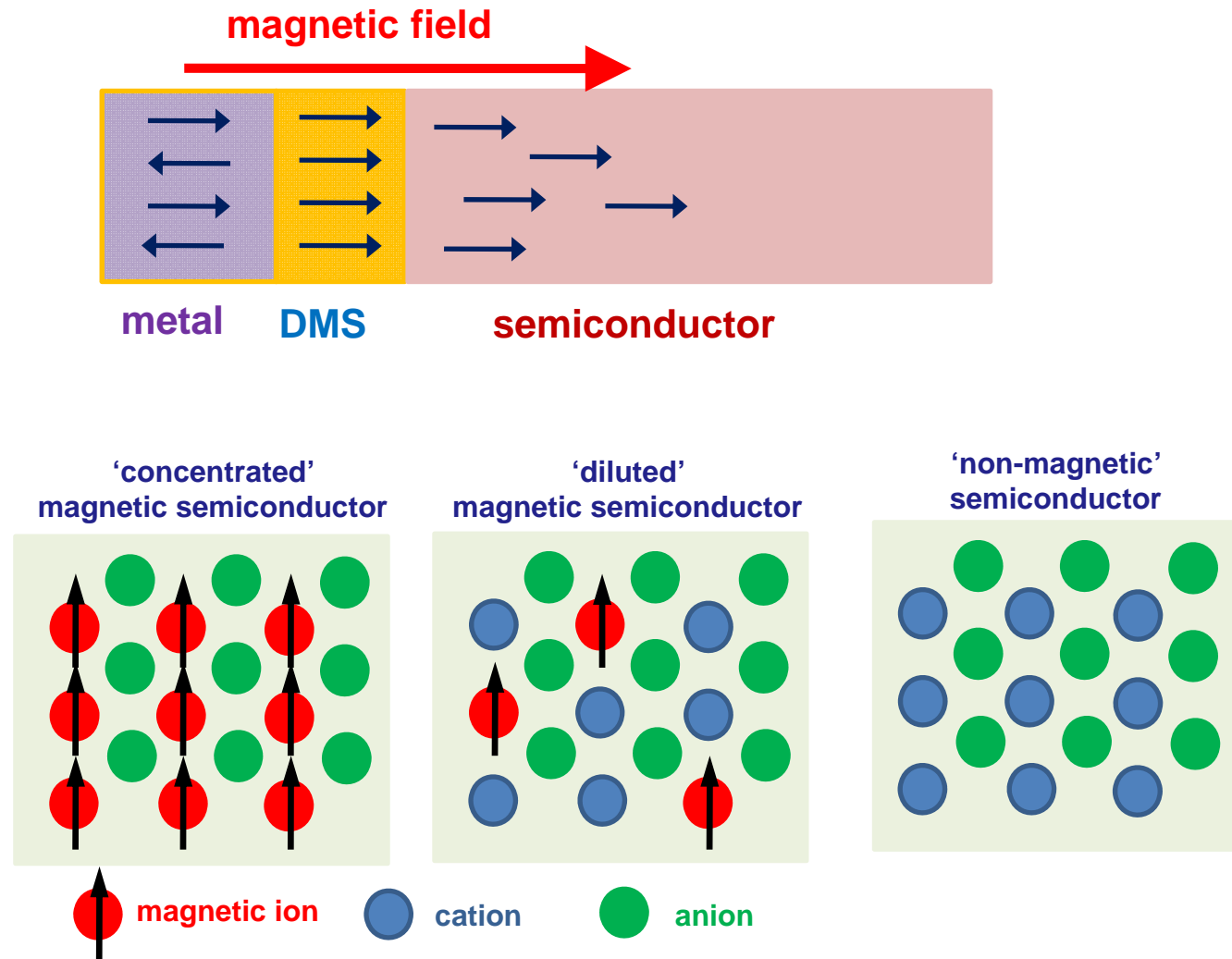
sample scanning stage



Space resolution: 200 nm ~ 1 um  
(nano-ARPES: 50 – 100 nm)  
X-ray photon energy: 100 – 1100 eV  
Surface sensitive (< 2 nm)  
UHV environment, in-situ experiments

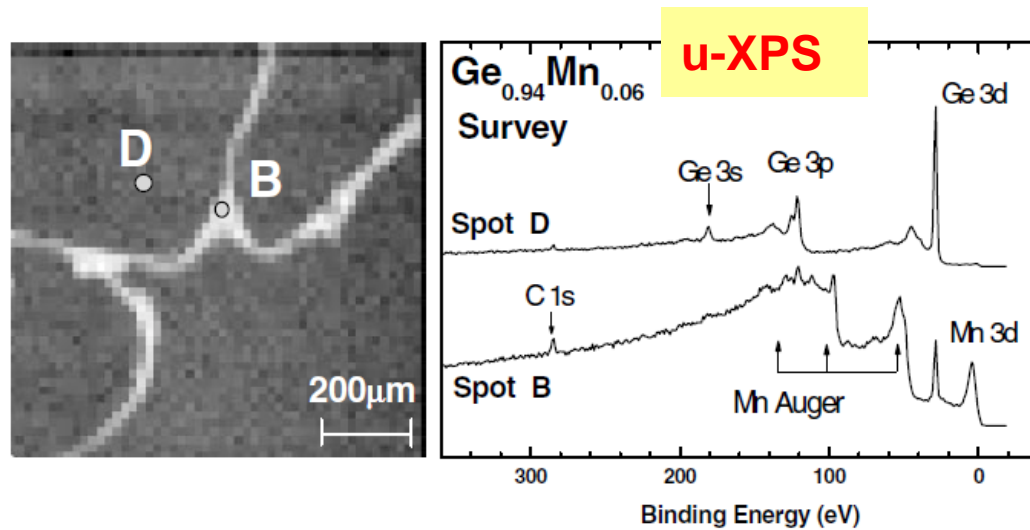
8A1 beamline, PLS

### Injection of spin-oriented electrons into semiconductor



## Observation of Mn rich phase in $\text{Ge}_{0.94}\text{Mn}_{0.06}$ DMS

DMS:  $\text{Ge}(0.94)\text{Mn}(0.06)$

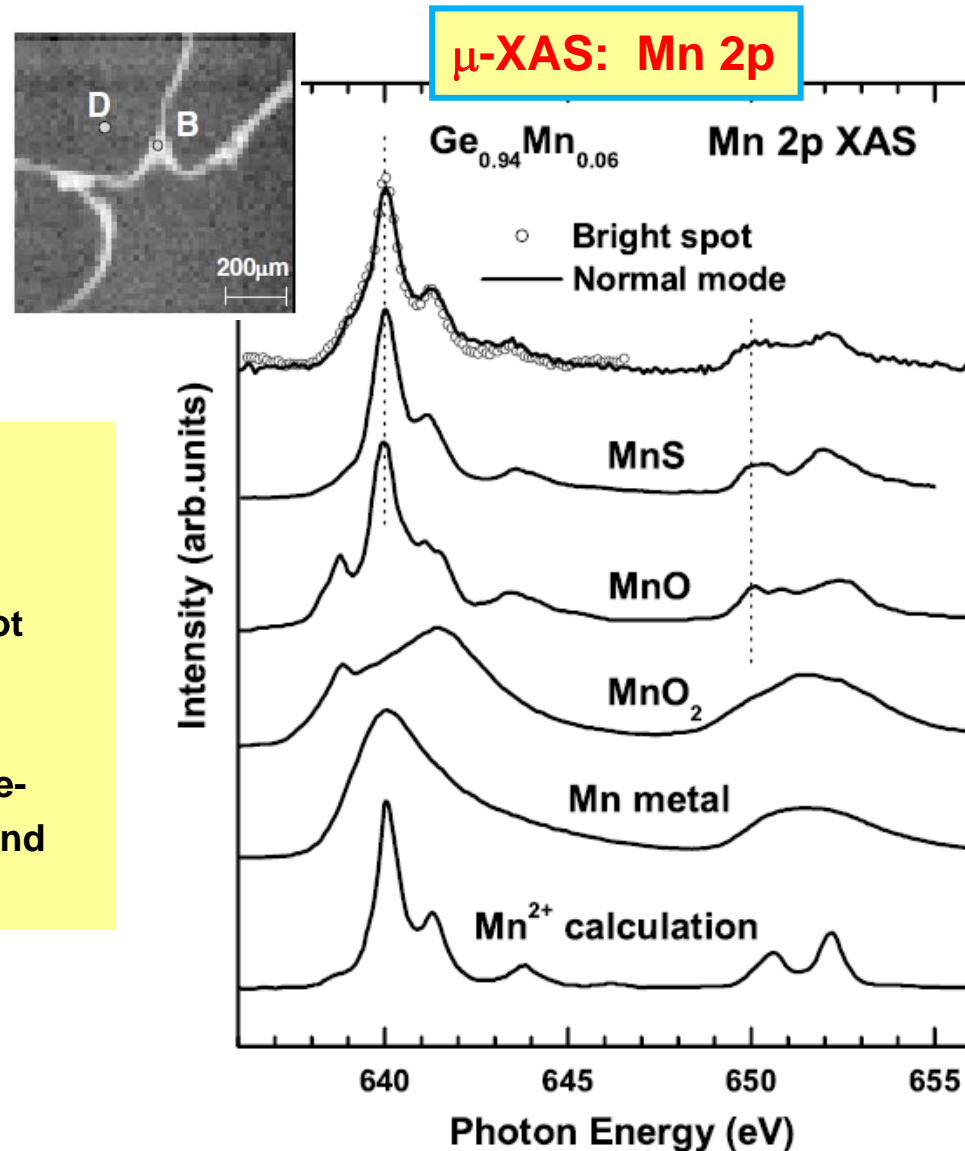


Observation of chemical phase separation into Mn rich and Mn depleted phases.

Similar phase separation of Cr and Fe -rich and -depleted phases were observed in the  $\text{Ge}(0.99)\text{Cr}(0.01)$  and  $\text{Ge}(1-x)\text{Fe}(x)$  DMS materials.

J. S. Kang, et al. , Phys. Rev. Lett. 94, 147202 (2005)

## Observation of Mn rich phase in $\text{Ge}_{0.94}\text{Mn}_{0.06}$ DMS



$\mu$ -XAS from the stripe-shaped microstructure.

→ Valence state of Mn ions:  $\text{Mn}^{2+}$ , not  $\text{Mn}^{4+}$ , nor  $\text{Mn}^0$  (metallic cluster).

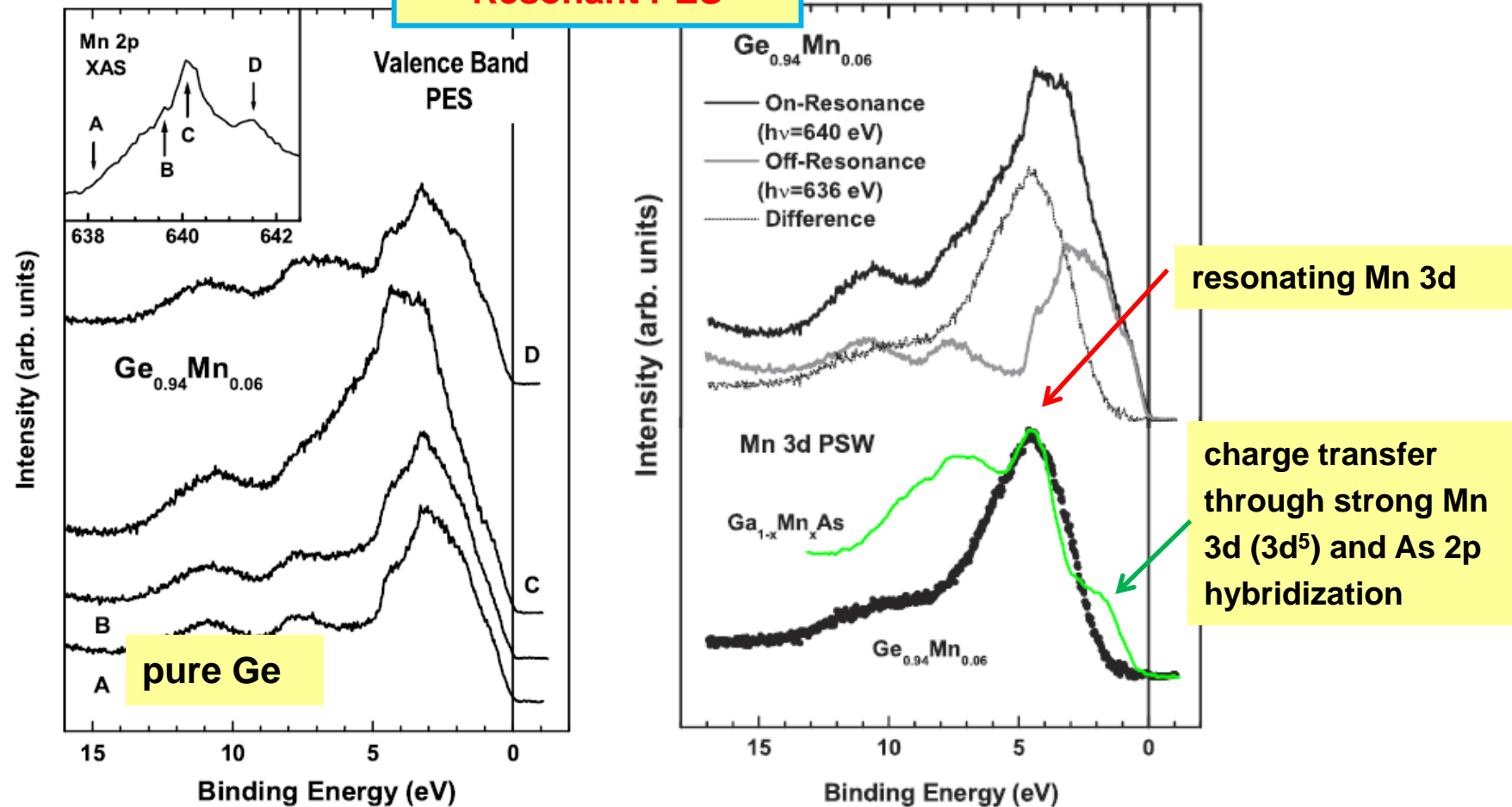
Theory expects that the Mn-rich stripe-shaped microstructure has divalent and high spin Mn ( $3d^5$ , total spin  $S = 5/2$ ).





## Observation of Mn rich phase in $\text{Ge}_{0.94}\text{Mn}_{0.06}$ DMS

### Resonant PES

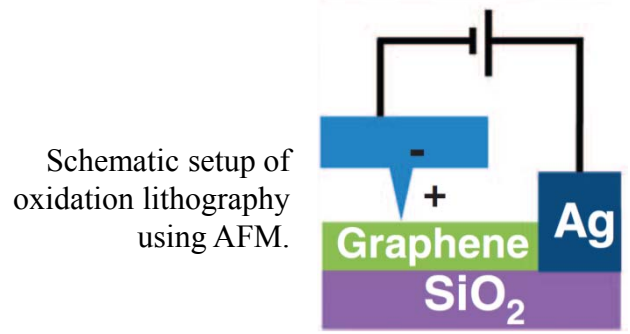


Mn 3d are located well below the  $E_F$ , and occupy the deep levels.  
 Confirms the  $\text{Mn}^{2+}$  valence state. Hybridization with Ge 2p is weaker.

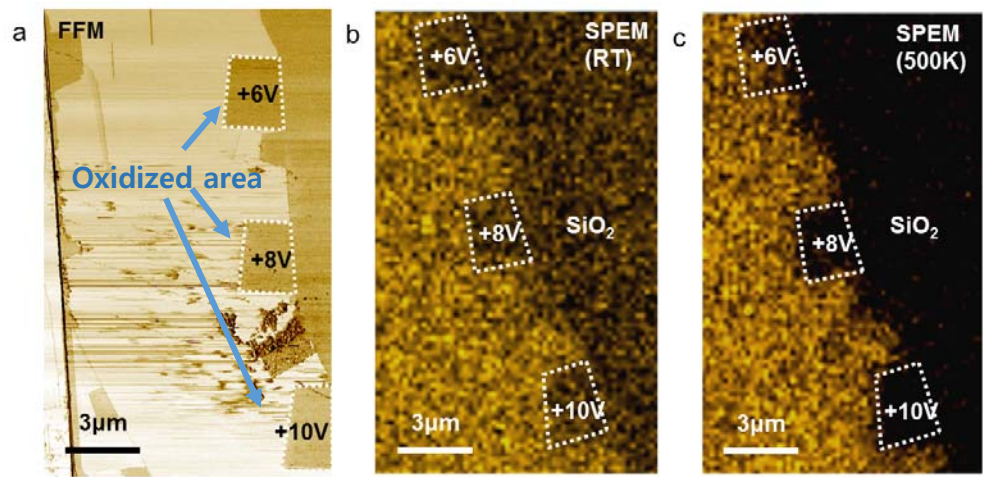
**The Mn-rich microstructure may explain the ferromagnetism in the  $\text{Ge}(0.94)\text{Mn}(0.06)$  DMS.**

## 2.3 SPEM application: control of graphene functionalization...

Electrical control of nanoscale functionalization in graphene by the scanning probe technique...

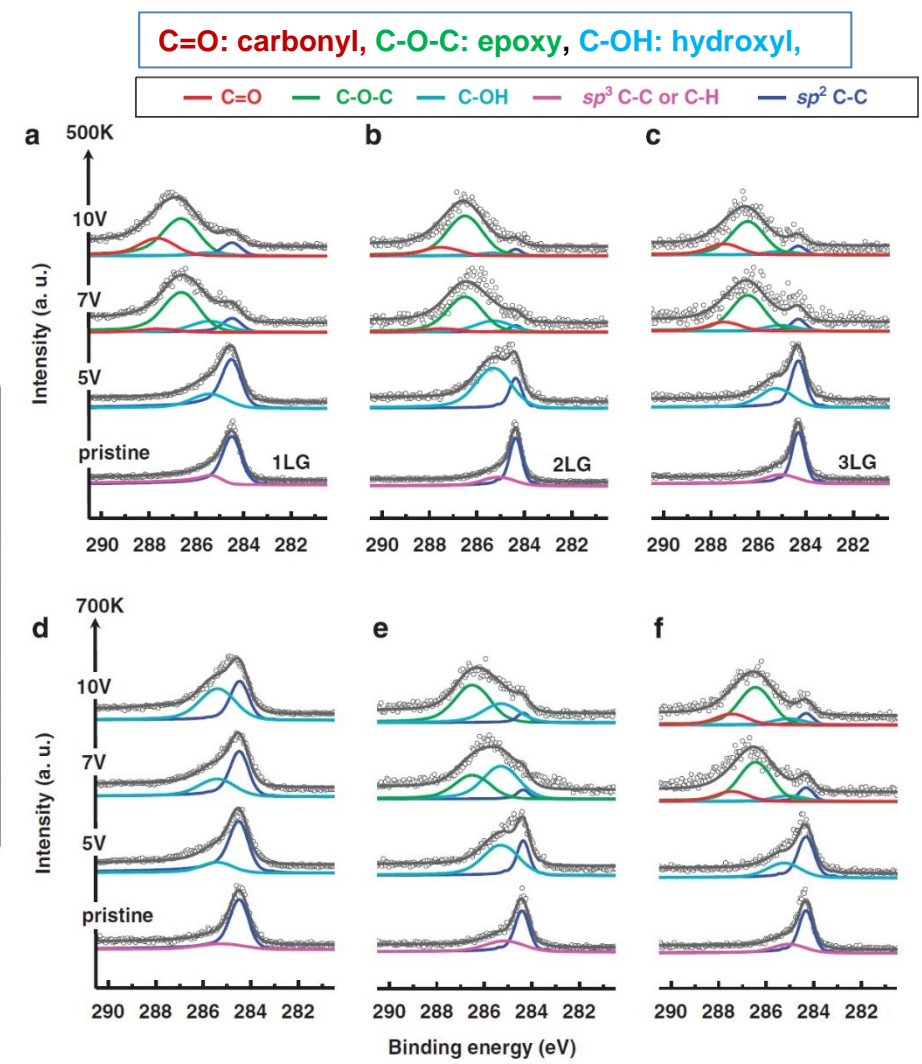


Effect of pre-annealing treatment (carbon contaminants removal)



FFM (a) and C1s@284.5eV SPEM (b and c) images of pristine and oxidized graphene on SiO<sub>2</sub> substrate before (a and b) and after (c) pre-annealing treatment at 500 K.

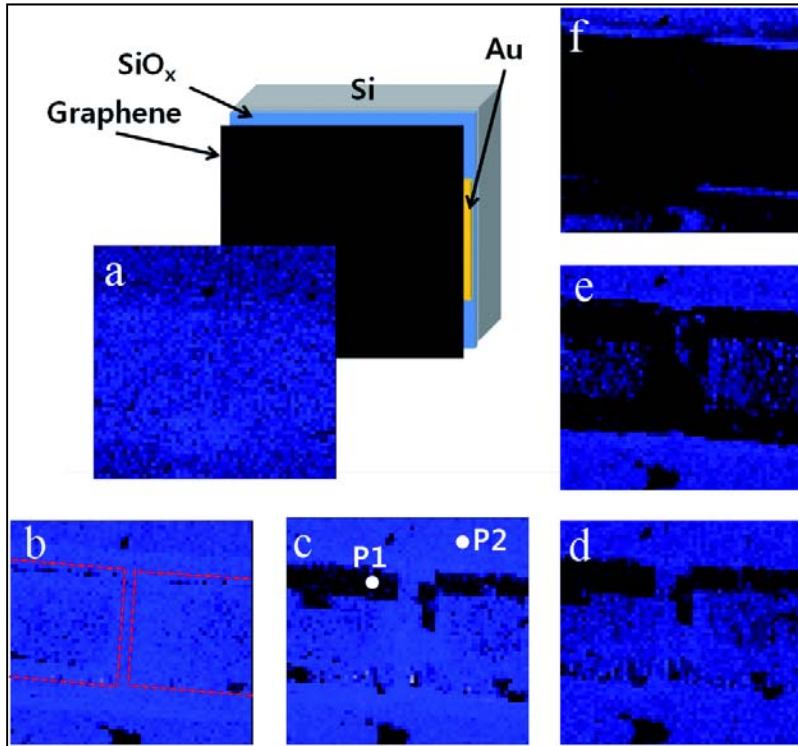
I. -S. Byun et al., NPG Asia Materials (2014) 6, e102



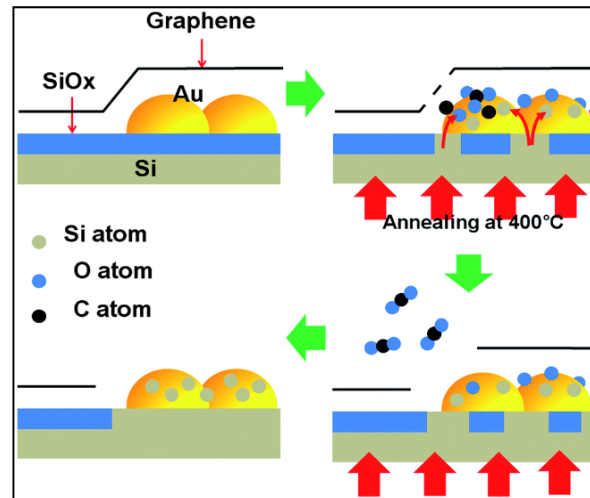
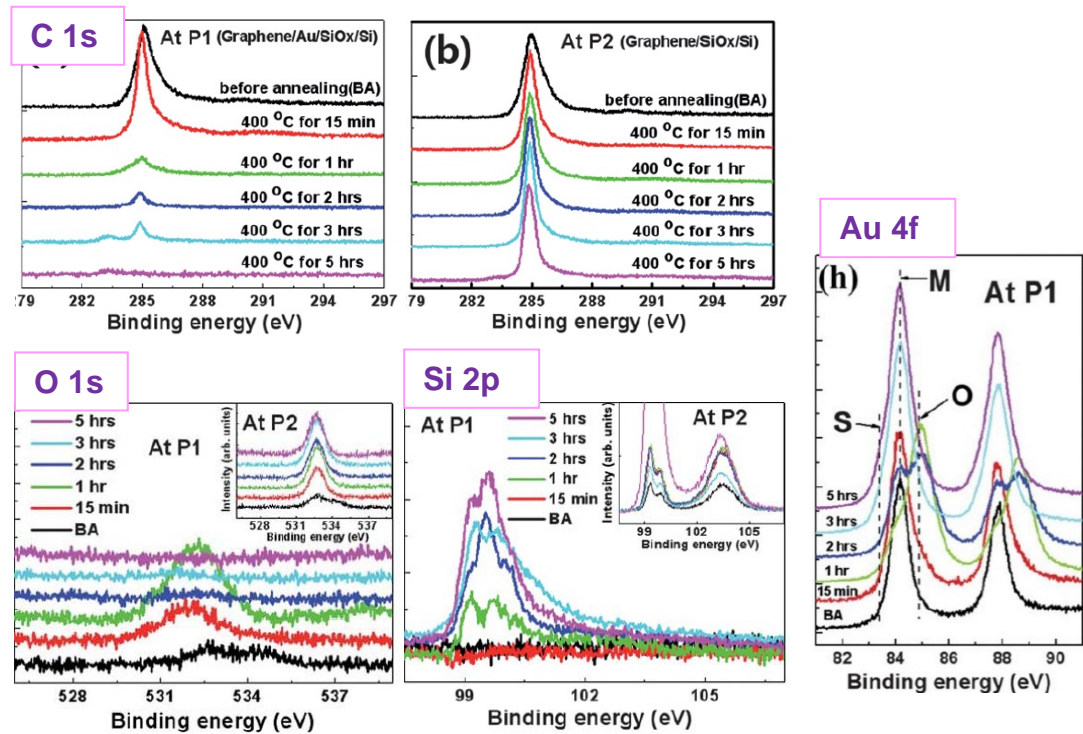
# 2.3 SPEM application: selective catalytic burning of graphene by SiO<sub>x</sub> layer depletion...

K. -J. Lee et al., *Nanoscale* 6, 1474-1479 (2014)

## C 1s SPEM images (600 μm x 600 μm)



(a) SPEM image (600 × 600 μm<sup>2</sup>), contrasted with the C 1s peak intensities, of the sample before annealing. Annealing time dependence of SPEM images (600 × 600 μm<sup>2</sup>), contrasted with C 1s peaks, at 400 °C: (b) 15 min, (c) 1 h, (d) 2 h, (e) 3 h, and (f) 5 h. Local points P1 and P2 indicate the graphene/Au/SiO<sub>x</sub>/Si and graphene/SiO<sub>x</sub>/Si surfaces, respectively.

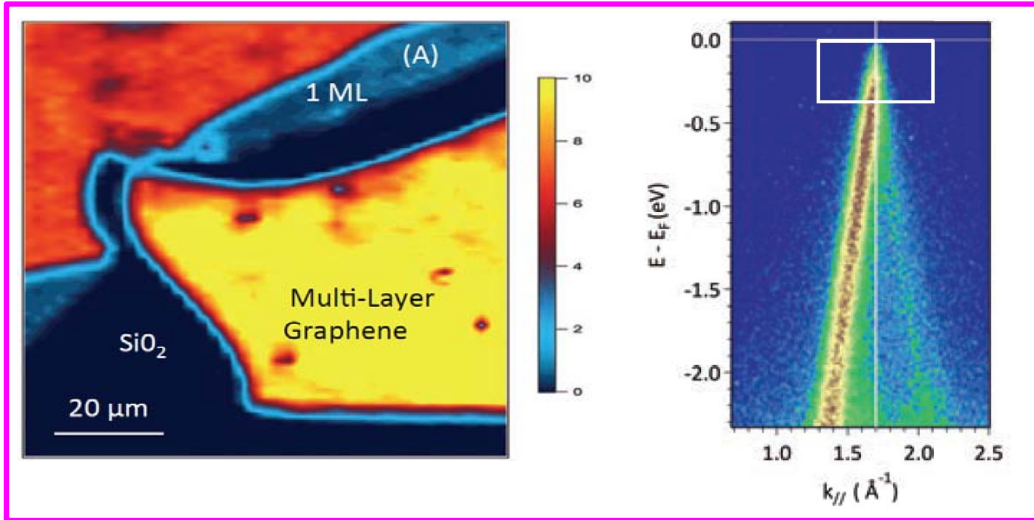


Schematic diagram of the catalytic oxidation process

Thermal dissociation of SiO<sub>x</sub>:  
 400 C / 737 °C  
 Contact resistivity:  
 26.17 / 1.58 × 10<sup>-4</sup> Ohm cm<sup>2</sup>



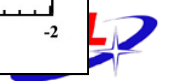
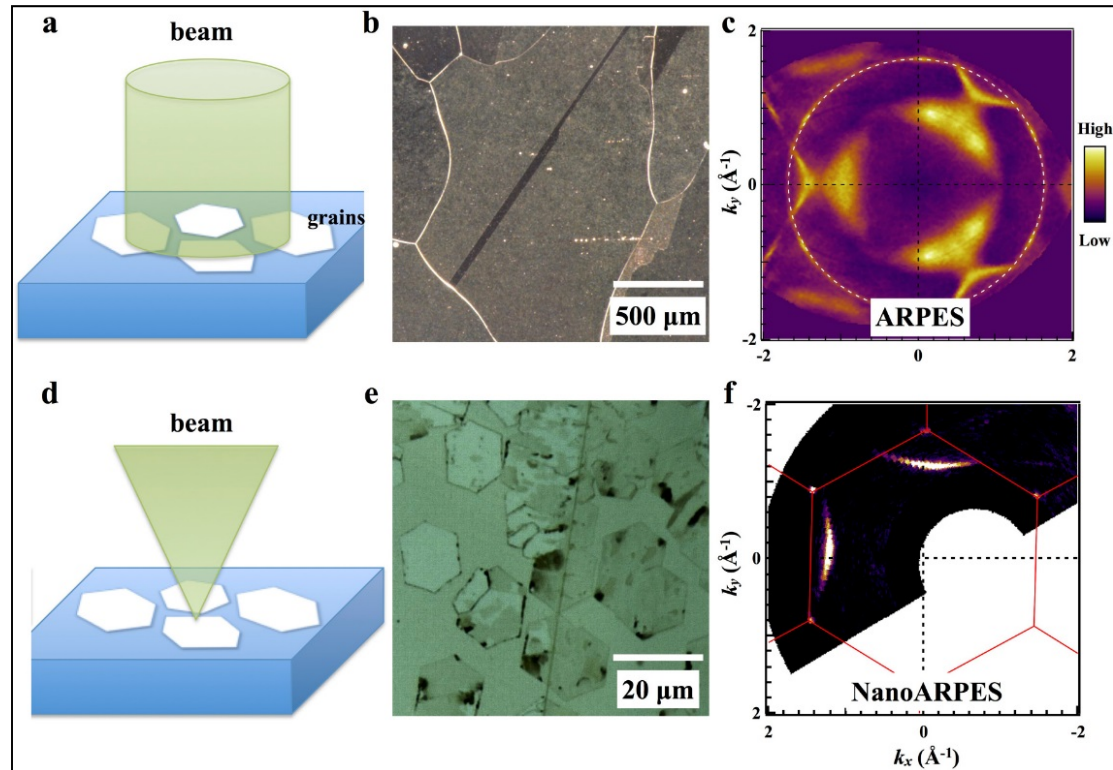
2.3 SPEM application: graphene layers (Nano-ARPES)



SOLEIL

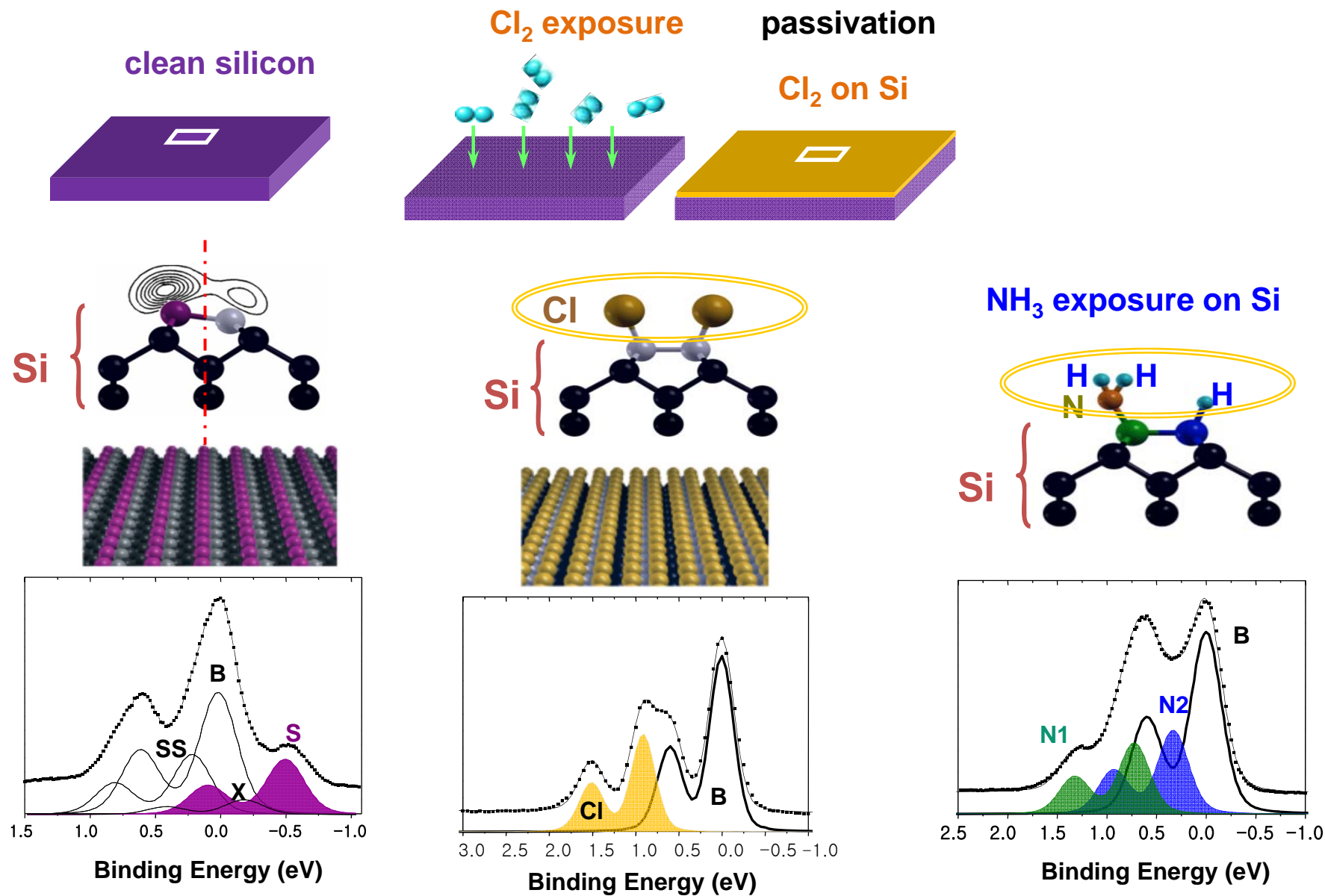
Limitations of traditional ARPES and the necessity of NanoARPES to study the intrinsic electronic structure of CVD graphene/Cu. (a) and (d) Scale difference of beam size and crystalline size in ARPES measurement. (b) and (e) Optical images of polycrystalline CVD graphene/Cu at different scales. (c) ARPES Fermi surface of polycrystalline CVD graphene/Cu as shown in (b) with a beam spot much bigger than the graphene domain size. The white dashed circle indicates the polycrystalline graphene FS. (f) NanoARPES FS from one single CVD graphene grain on Cu foil as shown in (e). Red hexagons indicate the Brillouin zone of graphene.

**C. Chen, et al, J. Phys.: Cond. Matt. (2017)**

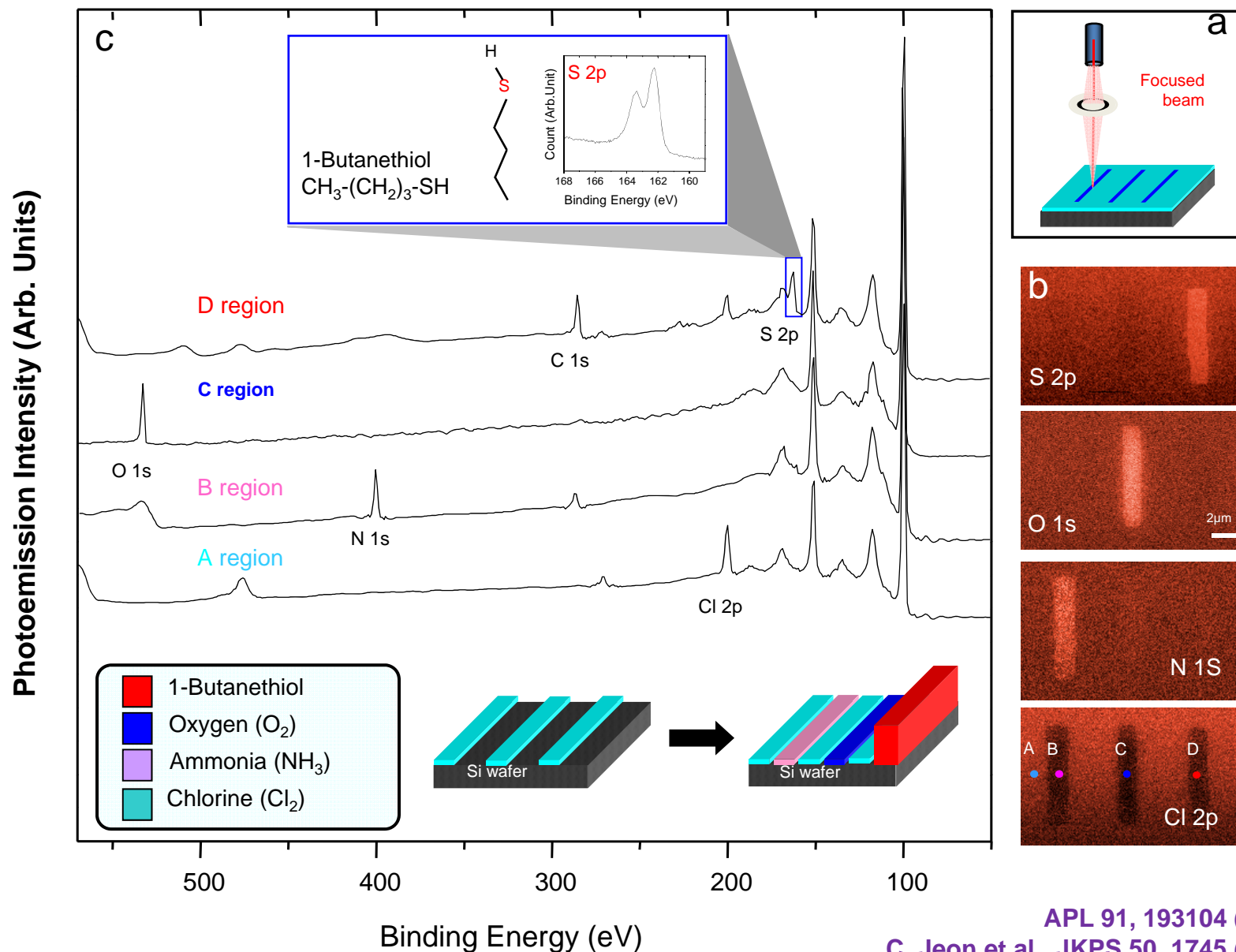




## 2.3 SPEM application: Atomic/molecular layer patterning



## 2.3 SPEM application: Atomic/molecular layer patterning



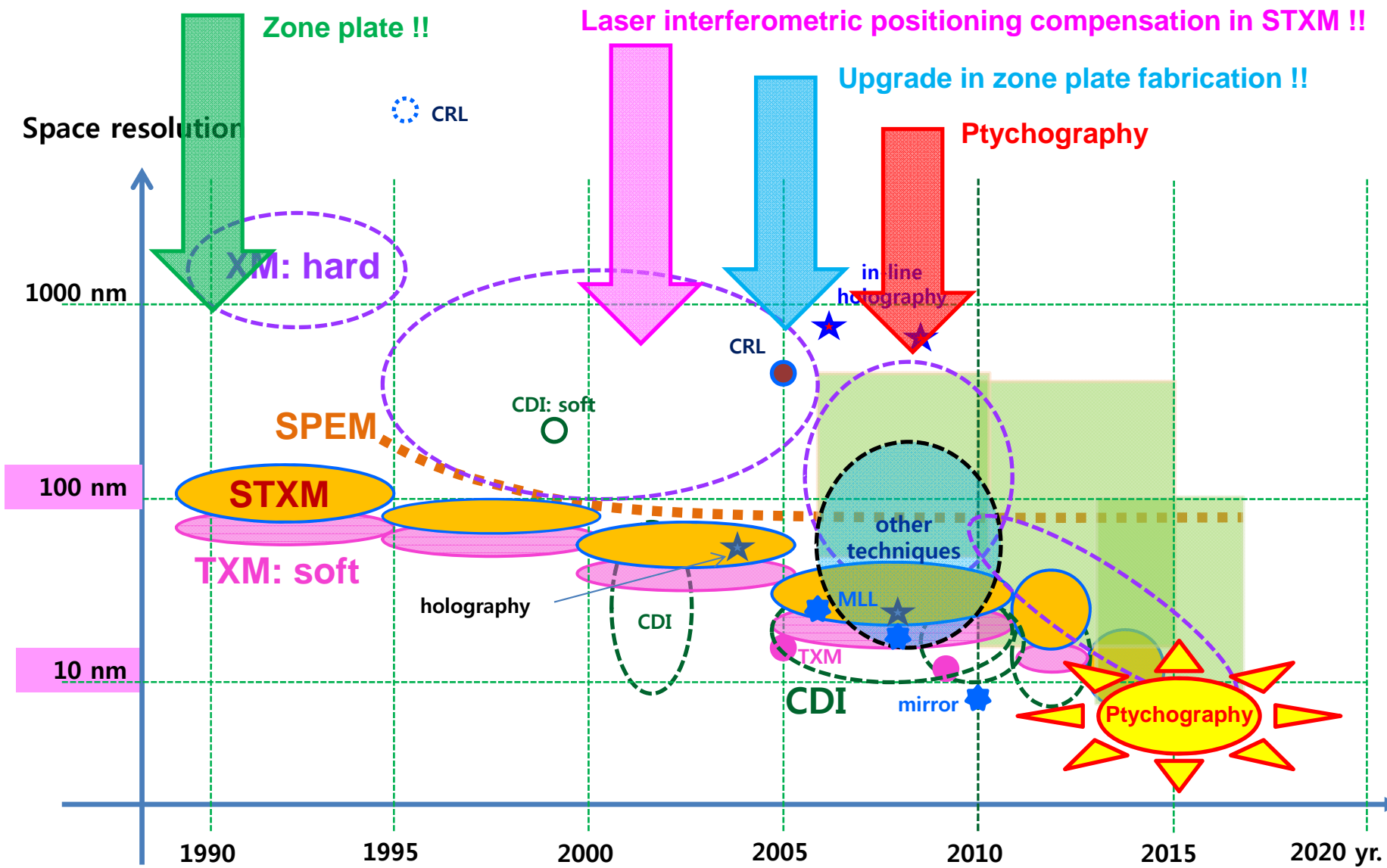
APL 91, 193104 (2007)

C. Jeon et al., JKPS 50, 1745 (2007)

S. W. Moon et al., Adv. Mat. 19, 1321 (2007)



### 3. Discussion: Evolution of space resolution of x-ray nanoscopy ..



### 3. Discussion: application fields ...

- Nano particles: catalytic reaction, ...
- Nano bio materials: nano ptl's influence on cell, ...
- Bio, medical application
- Environmental materials
- Polymers
- Thin film devices (in-situ degradation)
- Secondary ion battery materials
- Magnetic materials
- Geo- and extraterrestrial- materials
- ...

- in-situ experiments (annealing, biasing, magnetic field application, time resolved experiment, ...) are feasible
- samples in solution environment
- soft materials (less radiation damage due to higher efficiency in contrast)
- polarization sensitive materials
- from high vacuum to atmospheric pressure environment
- ...



### 3. Discussion: soft x-ray microscopes... (last changed: 31-Jan-2012)

Originally published in Handbook of Nanoscopy, Vol 2 (2012)

Courtesy of Prof. A. Hitchcock

Type	Facility	name	City	Country	Source	E-range (eV)	Status
TXM	Alba	Mistral	Barcelona	Spain	BM	270-2600	construction
TXM	ALS	XM1	Berkeley	USA	BM	250-900	operating
TXM	ALS	XM2 (NCXT)	Berkeley	USA	BM	250-6000	operating
TXM	Astrid	XRM	Aarhus	Denmark	BM	500	operating
TXM	Bessy	U41-TXM	Berlin	Germany	Und-L	250 - 600	operating
TXM	Diamond	B24 cryo-TXM	Harwell	UK	BM	250-2500	construction
TXM	NSRL	TXM	Hefei	China	BM	500	operating
TXM	Ritsumeikan	BL12	Kyoto	Japan	BM	500	operating
STXM	ALS	5.3.2.2	Berkeley	USA	BM	250-750	operating
STXM	ALS	5.3.2.1	Berkeley	USA	BM	250-2500	commissioning
STXM	ALS	11.0.2	Berkeley	USA	EPU	100-2000	operating
STXM	Bessy	old-STXM	Berlin	Germany	BM	250-600	decommissioned
STXM	Bessy	MAXYMUS	Berlin	Germany	EPU	250 - 1500	operating
STXM	CLS	10ID1	Saskatoon	Canada	EPU	130-2500	operating
STXM	Diamond	I08	Harwell	UK	EPU	250-2500	construction
(S)TXM	Elettra	Twin-mic	Trieste	Italy	Und-L	250 - 2000	operating
STXM	UVSOR	BL4U	Okazaki	Japan	Und-L	50-800	construction
STXM	NLSL	X1A (2)	Upton	USA	Und-L	250-1000	decommissioned
STXM	PLS	nanoscopy	Pohang	Korea	EPU	100 - 2000	construction
STXM	SLS	PolLux	Villigen	Switzerland	BM	250-750	operating
STXM	SLS	NanoXAS	Villigen	Switzerland	BM	250-750	commissioning
STXM	Soleil	Hermes	Saint-Aubin	France	EPU	250 - 1500	construction
STXM	SSRF	SXS	Shanghai	China	EPU	200 - 2000	operating
STXM	SSRL	13-1	Stanford	USA	EPU	250 - 1000	operating
SPEM	ALS	BL 7.0	Berkeley	USA	Und-L	90-1300	decommissioned
SPEM	ALS	Maestro	Berkeley	USA	EPU	90-1300	commissioning
SPEM	Elettra	BL 2.2 L	Trieste	Italy	Und-L	200-1400	operating
SPEM	Elettra	BL 3.2 L	Trieste	Italy	Und-L	27, 95	construction
SPEM	MAX-lab	BL 31	Lund	Sweden	Und-L	15-150	operating
SPEM	NSRRC	BL09A1	Hinschu	Taiwan	Und	60-1500	operating
SPEM	PAL	8A1	Pohang	Korea	Und	20 - 2000	operating
SPEM	Soleil	Antares	Saint-Aubin	France	EPU	50 - 1500	commissioning